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Synthesis, Biological Valuation and Molecular Docking Analysis of New 5-Benzylidene Bis-Rhodanine Derivatives

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ABSTRACT

The synthesis of 5-benzylidene bis-rhodanine derivatives are reported from bis-rhodanine (III) and different aromatic aldehydes (IV) via Knoevenagel condensation reactions. All the Derivatives (V) and (Va-m) were deep-rooted by NMR spectroscopic techniques and elemental analysis. The antiproliferative study of the compounds on HeLa human cervical cancer cell line, K562 leukemic cell line and MDAMB231 breast cancer cell line were performed by MTT assay. Docking studies were carried out against the protein HPV 16E2 present in the HeLa cell line. It show good docking scores. The results indicate that the bis-rhodanine derivatives could serve as potential molecules for the development of new anticancer agents.

Keywords: 5-benzylidene bis-rhodanine, Knoevenagel condensation, Cytotoxic studies, docking studies.

INTRODUCTION

Heterocyclic compounds are a major part of synthetic therapeutic chemistry. They offer a high level of structure variety and have demonstrated to be generally utilized as therapeutic agents. They are distributed widely as a natural product. Heterocyclic compounds have great potential and the most encouraging molecule for the discovery of new drugs. Ever since the introduction of Ciglitazone and Epalrestat for the treatment of diabetes, rhodanine has attracted medicinal chemists showing favourable biological properties.^{1,2,3} The chemical derivatization of rhodanine gives rise to molecules with a broad range of therapeutic activities like antiviral^{4,5}, antifungal^{6,7,8}, antibacterial⁹⁻¹⁶, antitumor¹⁷, aldose reductase inhibitors¹⁸⁻²⁰ and anti-inflammatory activities. Rhodanine has also been reported as inhibitors of uridine diphosphate-N-acetyl/L-alanine ligase²¹, hepatitis C virus protease inhibitors²² and also inhibitors of cancer cell migration²³⁻²⁶. From the laboratory, it has been reported 5-benzylidene-3-ethyl rhodanine and 5-isopropylidene-3-ethyl rhodanine induced cytotoxicity in a time and concentration-dependent manner with an IC₅₀ value of <10 μ M^{27,28}. Recently focus has been given on the mixture of bis-heterocyclic compounds, which showed various biological activities²⁹⁻³². As part of researcher's growing interest in synthesizing bis-rhodanine³³. we describe herein an easy and

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inexpensive synthetic route for the synthesis of a set bis-rhodanine to find novel and more potent anticancer agent.

Experimental procedure

General procedure for the synthesis of Bisrhodanine(III)

1.2-diamino propane (0.01 mol) was dissolved in water and while stirring then sodium Hydroxide (0.04 mol) was added. The solution was cooled at 0°C, the solution of carbon disulfide (0.02 mol) was mixed into it and kept for stirring for about 4 hours. Then the water solution of sodium chloroacetate (0.02 mol) was mixed with it and stirred for 3 hours. Subsequently, the solution of HCI was added and refluxed for 2-3 hours. While cooling a precipitate appeared and then it was filtered and dried to the corresponding product(III).

3,3'-(propane-1,2-diyl)bis(2-thioxothiazolidin-4one): (III)

Red colour solid, 58%, m.p. 232°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.5 (3H, d, J=6.8 Hz, H-1"), 4.5 (1H, m, H-2"), 4.05 (2H, m, H-3"), 4.15(2H, m, H-5), 4.20 (2H, m, H-5').¹³C-NMR (100 MHz, CDCl₃): δ 203.47 and 204.41 (C=S), 174.62 and 174.54 (C=O), 51.02 (N–CH), 44.9 (N-CH2) 35.58 and 34.28 (S–CH2), 14.46 (CH₂).

Synthesis of 5-benzylidene bis-rhodanine derivatives(V) & (Va-m)

The mixture of bis-rhodanine (III) (0.002 mol) and substituted aromatic aldehyde (IV) and (IVa-n) (0.004 mol) were refluxed in glacial acetic acid (0.004 mol) and sodium acetate (0.004 mol) for 4-6 hours. The resultant mixture on cooling yielded a precipitate which was filtered, washed, dried and recrystallised (ethanol) to afford the corresponding product (V) & (Va-m).

Compound V: Yellow solid, 58%, m.p. 298°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.64 (3H, d, J=7.5 Hz, H-1"), 4.75 (1H, m, H-2"), 3.84 (2H, m, H-3"), 7.78 and 7.84 (2H, S, H-6 and 6'), 7.98-8.21 (10H, m, H-8 to 12 and 8'-12'). ¹³C-NMR (100 MHz, CDCl₃): δ 203.18 and 201.80 (C=S), 174.50 and 174.04 (C=O), 115.27 and 114.66 (S-C), 150.13 and 148.83 (C-6 & 6' benzylic carbon), 125.16 to 133.7 (Aromatic carbon), 44.21 (N-CH2), 50.70 (N–CH), 14.77 (CH₃); Found: C-57.24, H-3.77, N-5.79, O-6.62, S-26.56.

Compound Va: Yellow solid, 63%, m.p. 296°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.62 (3H, d, J=6.7 Hz, H-1"), 4.65 (1H, m, H-2"), 3.94 (2H, m, H-3"), 3.66 (3H, S, OCH₃) 7.76 and 7.81 (2H, S, H-6 and 6'), 7.92-8.24 (8H, d, J=7.8 Hz, H-8 to 12 and 8'-12'). ¹³C-NMR (100 MHz, CDCl₃): δ 203.18 and 200.85 (C=S), 174.23 and 174.44 (C=O), 114.28 and 114.73 (S-C), 150.24 and 148.78 (C-6 & 6' benzylic carbon), 125.38 to 134.6 (Aromatic carbon), 56.3 (OCH₃), 45.54 (N-CH2), 50.10 (N–CH), 14.13 (CH₃); Found:C-55.34, H-4.08, N-5.17, O-11.78, S-23.62.

Compound Vb: Yellow solid, 66%, m.p. 296°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.56 (3H, d, J=6.8 Hz, H-1"), 4.81 (1H, m, H-2"), 3.77 (2H, m, H-3"), 7.58 and 7.67 (2H, S, H-6 and 6'), 7.91-8.29 (8H, d, J=7.7 Hz, H-8,9,11,12 and 8',9',11',12'). ¹³C-NMR (100 MHz, CDCl₃): δ 203.28 and 201.14 (C=S), 172.42 and 172.33 (C=O), 116.28 and 115.77 (S-C), 151.12 and 149.64 (C-6 & 6' benzylic carbon), 125.35 to 134.9 (Aromatic carbon), 45.14 (N-CH2), 51.77 (N–CH), 14.91 (CH₃); Found: C-50.07, H-2.93, Cl-12.85, N-5.07,O-5.82, S-23.24.

Compound Vc: Yellow solid, 65%, m.p. 294°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.62 (3H, d, J=6.7 Hz, H-1"), 4.71 (1H, m, H-2"), 3.92 (2H, m, H-3"), 7.88 and 7.97 (2H, S, H-6 and 6'), 7.92-8.35 (8H, d, J=7.5 Hz, H-8,9,11,12 and 8',9,11',12'). ¹³C-NMR (100 MHz, CDCl₃): δ 204.38 and 203.85 (C=S), 174.02 and 174.21 (C=O), 115.11 and 114.63 (S-C), 151.02 and 149.82 (C-6 & 6' benzylic carbon), 125.36 to 134.7 (Aromatic carbon), 44.25 (N-CH2), 50.70 (N-CH), 15.05 (CH₃); Found: C-43.15, H-2.54,Br-24.98, N-4.34, O-5.03, S-20.05.

Compound Vd: Yellow solid, 65%, m.p. 290°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.65 (3H, d, J=6.8 Hz, H-1"), 4.85 (1H, m, H-2"), 3.85 (2H, m, H-3"), 7.79 and 7.83 (2H, S, H-6 and 6'), 7.96-8.25 (8H, d, J=7.5 Hz, H-8,9,11,12 and 8',9',11',12'). ¹³C-NMR (100 MHz, CDCl₃): δ 203.38 and 201.85 (C=S), 174.52 and 174.34 (C=O), 115.18 and 114.83 (S-C), 151.14 and 149.94 (C-6 & 6' benzylic carbon), 125.38 to 134.7 (Aromatic carbon), 45.15 (N-CH2), 51.80 (N–CH), 14.85 (CH₃); Found: C-48.25, H-2.83, N-9.77, O-16.77, S-22.42.

Compound Ve: Yellow solid, 62%, m.p. 291°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.64 (3H, d, J=6.7 Hz, H-1"), 4.84 (1H, m, H-2"), 3.76 (2H, m,

H-3"), 2.51 (3H, S, CH₃) 7.68 and 7.79 (2H, S, H-6 and 6'), 7.94-8.31 (8H, d, J=7.8 Hz, H-8,9,11,12 and 8',9,11',12'). ¹³C-NMR (100 MHz, CDCl₃): δ 200.11 and 200.92 (C=S), 172.54 and 172.96 (C=O), 115.88 and 114.97 (S-C), 151.32 and 150.27 (C-6 & 6' benzylic carbon), 125.33 to 134.60 (Aromatic carbon), 43.26 (N-CH2), 50.54 (N–CH), 28.4 (phenyl methyl), 14.05 (CH₃); Found: C-58.78, H-4.33, N-5.48, O-6.28, S-25.12.

Compound Vf: Yellow solid, 63%, m.p. 302° C, ¹H-NMR (400 MHz, CDCl₃): δ 1.46 (3H, d, J=6.7 Hz, H-1"), 4.63 (1H, m, H-2"), 3.77 (2H, m, H-3"), 7.72 and 7.85 (2H, S, H-6 and 6'), 7.88-8.33 (8H, d, J=7.5 Hz, H-8,9,11,12 and 8',9',11',12'). ¹³C-NMR (100 MHz, CDCl₃): δ 200.38 and 200.76 (C=S), 174.33 and 174.01 (C=O), 115.17 and 114.81 (S-C), 151.11 and 149.90 (C-6 & 6' benzylic carbon), 125.30 to 134.6 (Aromatic carbon), 44.18 (N-CH2), 51.86 (N–CH), 14.81 (CH₃); Found: C-53.27, H-3.12,F-7.34, N-5.42, O-6.18, S-24.72.

Compound Vg: Yellow solid, 63%, m.p. 288°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.56 (3H, d, J=6.8 Hz, H-1"), 4.80 (1H, m, H-2"), 3.63 (2H, m, H-3"), 5.44 (1H, S, OH) 7.73 and 7.81 (2H, S, H-6 and 6'), 7.95-8.26 (8H, d, J=7.8 Hz, H-8,9,11,12 and 8',9',11',12'). ¹³C-NMR (100 MHz, CDCl₃): δ 203.11 and 203.85 (C=S), 170. 42 and 171.03 (C=O), 114.98 and 114.21 (S-C), 151.03 and 150.14 (C-6 & 6' benzylic carbon), 125.35 to 135.5 (Aromatic carbon), 44.15 (N-CH2), 50.66 (N–CH), 14.79 (CH₃); Found: C-53.68, H-3.53, N-5.44,O-12.43, S-24.92.

Compound Vh: Yellow solid, 59%, m.p. 287°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.63 (3H, d, J=6.8 Hz, H-1"), 4.72 (1H, m, H-2"), 3.81 (2H, m, H-3"), 7.70 and 7.81 (2H, S, H-6 and 6'), 7.95-8.29 (8H, d, J=7.6 Hz, H-8 to 10,12 and 8-10',12'). ¹³C-NMR (100 MHz, CDCl₃): δ 203.30 and 202.66 (C=S), 170.51 and 171.24 (C=O), 115.33 and 114.78 (S-C), 151.23 and 149.99 (C-6 & 6' benzylic carbon), 125.30 to 134.65 (Aromatic carbon), 42.11 (N-CH2), 51.01 (N–CH), 14.77 (CH₃); Found: C-50.08, H-2.93,Cl-12.85, N-5.09, O-5.80, S-23.25.

Compound Vi: Yellow solid, 64%, m.p. 287°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.64 (3H, d, J=6.8 Hz, H-1"), 4.86 (1H, m, H-2"), 3.86 (2H, m, H-3"), 7.74 and 7.80 (2H, S, H-6 and 6'), 7.99-8.21 (8H, d, J=7.7 Hz, H-8 to 10,12 and 8'-10',12').

¹³C-NMR (100 MHz, CDCl₃): δ 200.38 and 200.85 (C=S), 170.52 and 171.32 (C=O), 116.13 and 113.81 (S-C), 151.12 and 149.95 (C-6 & 6' benzylic carbon), 125.36 to 134.5 (Aromatic carbon), 45.19 (N-CH2), 51.85 (N-CH), 14.82 (CH₃); Found:C-43.12, H-2.53, Br-24.94, N-4.38, O-5.02, S-20.04.

Compound Vj: Yellow solid, 60%, m.p. 299°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.64 (3H, d, J=6.8 Hz, H-1"), 4.89 (1H, m, H-2"), 3.86 (2H, m, H-3"), 7.79 and 7.88 (2H, S, H-6 and 6'), 7.94-8.22 (8H, d, J=7.6 Hz, H-8 to 10,12 and 8'-10',12'). ¹³C-NMR (100 MHz, CDCl₃): δ 201.39 and 202.87 (C=S), 175.58 and 175.32 (C=O), 114.17 and 114.82 (S-C), 150.18 and 149.97 (C-6 & 6' benzylic carbon), 125.39 to 134.9 (Aromatic carbon), 45.18 (N-CH2), 51.85 (N-CH), 14.84 (CH₃); Found: C-53.22, H-3.11,F-7.36, N-5.43, O-6.19, S-24.71.

Compound Vk: Yellow solid, 60%, m.p. 289°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.66 (3H, d, J=6.8 Hz, H-1"), 4.87 (1H, m, H-2"), 3.89 (2H, m, H-3"), 7.74 and 7.81 (2H, S, H-6 and 6'), 7.95-8.27 (6H, d, J=7.5 Hz, H-8 to 10 and 8'-10'). ¹³C-NMR (100 MHz, CDCl₃): δ 202.39 and 201.83 (C=S), 174.51 and 174.34 (C=O), 115.17 and 114.82 (S-C), 151.13 and 149.98 (C-6 & 6' benzylic carbon), 125.39 to 134.5 (Aromatic carbon), 45.17 (N-CH2), 51.85 (N–CH), 14.88 (CH₃); Found: C-53.24, H-3.19, F-7.35, N-5.42, O-6.18, S-24.74.

Compound VI: Yellow solid, 58%, m.p. 295°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.68 (3H, d, J=6.8 Hz, H-1"), 4.88 (1H, m, H-2"), 3.84 (2H, m, H-3"), 7.78 and 7.81 (2H, S, H-6 and 6'), 7.97-8.28 (4H, S, H-8, 12 and 8',12'). ¹³C-NMR (100 MHz, CDCl₃): δ 203.39 and 201.84 (C=S), 174.51 and 174.35 (C=O), 115.19 and 114.85 (S-C), 151.13 and 149.95 (C-6 & 6' benzylic carbon), 125.39 to 134.8 (Aromatic carbon), 45.17 (N-CH2), 58.8 (OCH₃), 51.82 (N-CH), 14.83 (CH₃); Found: C-44.53, H-2.26, Cl-22.85, N-4.53, O-5.17, S-20.66.

Compound Vm: Yellow solid, 65%, m.p. 290°C, ¹H-NMR (400 MHz, CDCl₃): δ 1.61 (3H, d, J=6.8 Hz, H-1"), 4.80 (1H, m, H-2"), 3.85 (2H, m, H-3"), 7.79 and 7.83 (2H, S, H-6 and 6'), 7.96-8.25 (8H, d, J=7.6 Hz, H-8,10,11,12 and 8',10',11',12'). ¹³C-NMR (100 MHz, CDCl₃): δ 203.38 and 201.85 (C=S), 174.52 and 174.34 (C=O), 115.18 and 114.83 (S-C), 151.14 and 149.94 (C-6 & 6' benzylic carbon),

125.38 to 134.7 (Aromatic carbon), 45.15 (N-CH2), 51.80 (N–CH), 14.85 (CH $_3$); Found: C-48.25, H-2.83, N-9.77, O-16.77, S-22.42.

Anticancer activity Cell Line

The cervical cell line HeLa, leukemic cell line K562 and the cell line MDAMB231 for breast cancer were received from NCCS-National Center for Cell Science, Pune and grown.

MTT assay

MTT assay was followed by the standard protocol according to Rajendran Kumar *et al.*, The worth of IC_{50} was calculated with a nonlinear regression graph that strategized amid current inhibition of cells in one axis and concentration of logs oneother.

LDH assay

Lactate dehydrogenase release assay liberates, Lactate dehydrogenase (LDH) is a pointer of membrane integrity and henceforth cell injury. Following treatment with compound Vf (3.12, 6.25, 12.5, 25, 50 and 100 µg/mL) on HeLa, K562, MDAMB231 cancer cells for 48 h and it was conducted to measure the proclamation of LDH on the way to the media and measured using standard protocols (Moorthy et al., 2010). The intracellular LDH was determined by fast congelation and melting in liquid nitrogen after losing the cells. The proclamation of LDH was calculated at an absorption rate of 490 nm. The releasing proportion of LDH was measured for example (media LDH activity)/(Media LDH activity) X 100%. Outcomes are exposed as a proportion of LDH release that substrates switch standards from the preserved ones.

FACS analysis

Analysis of the FACS was performed to determine compound 1 had any effect on the development of cell cycles. After 48 h of 50 and 100 μ g/mL treatments, MDAMB231 cells stained with propidium iodide and endangered to FACS. The DMSO treated cells showed a regular pattern for the cell rotation. Cells were treated with compound Vf absorptions of 50 and 100 μ g/mL and incubated for further 24 hours. At room temperature, cells were gathered then centrifuged for 5 min at 2000rpm. The supernatant was carefully removed, preserving the cell pellet. The cell pellet washed through re-dangling in 2 mL of 1XPBS. With the same conditions, the washing was repeated for another time. The supernatant abandoned, with the pellet was retained. Cells set by resuspending in 300 μ L of Sheath fluid. Preceded by adding 1 mL of chilled 70% EtOH dropwise with constant gentle shaking, and adding another 1 mL again.

Docking studies

The mark protein nominated for the current research is HPV 16 E2 protein. The manufactured bis-rhodanine derivatives were practiced for molecular docking study.

Molecular Docking

The study of Docking under the default setting was done using Autodock with a binding pocket on the TAD. The result was analyzed using PYMOL (TM) by visualizing the protein-ligand and calculating parameters like hydrogen bonding amid protein and the ligand thus, determining scores are tabulated.

RESULT AND DISCUSSION

While doing the literature survey to the best of our knowledge, there is only one report available for the synthesis of bis-rhodanine using diamines, Carbon disulfide and dialkylacetylenedi carboxylates³². In the present work, diamines, Carbon disulfide, and chloroacetic acid for the synthesis of bis-rhodanine are used. To found the optimal reaction conditions for the integration of bis-rhodanine, it is decided to begin with the conventional method. In this method, one mole of diamine and two moles of carbon disulfide and chloroacetic acid are used. The dithioamide is formed initially by the reaction of carbon disulfide and diamine serves as a nucleophile and reacts further with chloroacetic acid followed by cyclization yield bis-rhodanine (III). This method have been reported earlier³³, but the mechanism was not explained adequately. Now the device is shown in Scheme 2.

New derivatives were synthesized by Knoevenagel condensation of bis-rhodanine(III) with different substituted aldehydes (IV) in the presence of sodium acetate and acetic acid to yield the products (V) and (Va-m). The newly synthesized 5-benzylidenebis-rhodanine derivatives were fully characterized by proton NMR, carbon NMR spectral data and elemental analysis.



In the ¹H-NMR spectra of a representative compound Ve in the upfield region the N-methylene protons resonated at δ 3.76 (2H, m, H-3"), the methyl group appeared as a doublet at δ 1.64 (3H, J = 6.7 Hz), and the corresponding methine proton appeared as a multiplet at δ 4.8 (1H, m, H-2"). The two singlets at δ 7.68 (1H, s, H-6) and 7.79 (1H, s, H-6') were attributed to the benzylic protons one on each side. The Phenyl protons appeared as a pair at δ 7.96 (4H, d, J = 7.8 Hz) and 8.15 (4H, d, J = 7.8 Hz). The CH₃ group attached to the phenyl ring resonated at 2.51 (3H, S, CH₃). The above data was complemented by its carbon NMR spectra. The pair of signals at δ 200.11 and 200.92 and δ 174.87 and 174.62 are due to thiocarbonyl (C=S) and carbonyl carbon

(C=O) of the rhodanine ring system respectively. The pair of signals at δ 115.88 and 114.97 are due to the =C-S group of the rhodanine system. The CH₃ carbon appeared at δ 14.05, and the CH₂ and CH carbons attached to the nitrogen resonated at 50.54 and 43.26. The signals at δ 151.32 and 150.27 are assigned to the benzylic carbons (C-6 & C-6'). The methyl carbon attached to the aromatic system appeared at δ 28.4. The bunches of signalsbetween 125.33 to 134.60 are attributed to the phenyl carbon atoms. All the other messages of the compound (V) and (Va-m) coincided with signs of compound(Ve).

Mechanism



Scheme 2. Synthesis of 5-benzylidene bis-rhodanine derivatives (V) and (Va-m)

m; 3-NO₂

Cytotoxic activity

The effect of compounds V and Va-m on the proliferation of HeLa, MDAMB231 and K562 cells were carried out using MTT assay. HeLa, MDAMB231, and K562 cells were added with 3.12, 6.25, 12.5, 25, 50 and 100 µM of compounds V and Va-m and after 48 h the assay was performed. IC₅₀ values were calculated and presented in the Table 1. The viability of the cells were affected at a moderate concentration in all the derivatives V and Va-m with HeLa, MDAMB231 and K562 cells and was more pronounced in the case of the compound Vf against MDAMB231 breast cancer cell lines. The compound V without any substituents should be an IC₅₀ value of 75.0, 85.4, and 72 μM concentration against HeLa, MDAMB231, and K562 cells. The IC_{50} value of all the other compounds Va-m which has substituents in the aromatic system has varied substantially from the IC₅₀ value of compound V against the three cell lines. This indicates that the substituents affect the proliferation activity against the three cell lines. If we divide these substituents into two groups like electron-withdrawing substituents (NO₂, F, Cl and Br) and electron-pumping substituents (-OCH_a, OH and $\rm CH_{_3})$ and consider the $\rm IC_{_{50}}$ value concerning HeLa cell lines, the activity in compounds with electronwithdrawing groups Vb, Vc, Vd, Vf showed lower IC₅₀ costs than the compounds with electron-donating substituents Va, Ve and Vg. As an example, the IC50 value of compound Va with an electron-donating substituent - OCH₃ is 81.2 µM.

In contrast, for the compound with electronwithdrawing substituent fluorine in Vf, the IC₅₀ value is 54 µM. The same trend is followed in MDAMB231 and K562 cells too. Further the compounds are divided into two groups depending upon the position of the substituent placed in the aromatic ring-like parasubstituted compounds Va to Vg and meta substituted compounds Vh, Vi, Vj, and Vm, the IC₅₀ value of parasubstituted compounds and relatively lower than the meta substituted compounds. The IC₅₀ value of Vf with a para-substituted fluorine group has an IC₅₀ value of 54 $\mu\text{M},$ whereas the corresponding compound with meta substituted fluorine group showed an IC₅₀ amount of 61 µM. Overall concerning all the three HeLa, MDAMB231, and K562 cell lines, the mixture Vf with a para-substituted fluorine atom performed well. It showed an IC₅₀ value of 54.0, 32.0, and 58.0 against HeLa, MDAMB231, and K562 cell lines, respectively. So compound Vf was taken for further studies.

Table 1: Inhibitory effect of compounds (V) and (Va-m)oncytotoxic activity by MTT assay

S No	Compound	I R	IC ₅₀ ,μM HeLa	MDAMB231	K562	Binding energy, kcal/mol
2	(V)	Н	75.0	85.0	72.0	-8.6
3	(Va)	p-OCH ₃	81.2	82.5	78.2	-9.5
4	(Vb)	p-Cl ຶ	77.0	78.0	79.0	-9.6
5	(Vc)	p-Br	55.5	65.2	60.5	-8.9
6	(Vd)	p-NO	66.5	76.5	70.5	-8.8
7	(Ve)	p-CH ₃	77.5	82.2	74.5	-9.3
8	(Vf)	p-F	54.0	32.0	58.0	-9.1
9	(Vg)	p-OH	66.5	69.5	69.5	-9.2
10	(Vh)	m-Cl	72.5	82.5	75.5	-8.8
11	(Vi)	m-Br	69.0	76.0	70.0	-8.7
12	(Vj)	m-F	61.0	44.0	64.0	-9.6
13	(Vk)	m,p-Cl	62.5	68.4	68.5	-9.5
14	(VI)	m,p-OCH ₃	65.5	69.3	67.5	-9.0
15	(Vm)	m-NO	80.0	83.0	82.0	-9.1
16	Cis platin		23.60	24.86	29.32	-

Lactate dehydrogenase discharge examine was performed to test the cell membrane harm incited by Compound Vf for this, the MDAMB231 cell line was refined with 10, 50, 100, and 250 μ M concentration of compound V_f and LDH discharged was estimated at 24, 48, and 72 hours. Predictable with the above outcomes a portion and timesubordinate increment in LDH discharge was monitered and confirming the cytotoxic capability of Compound Vf Fig. 1 was confirmed.



Fig. 1. LDH assay of compound Vf against MDAMB231 cell line

FACS analysis

Fluorescence-activated Cell Sorting investigation was performed to decide if compound V_f has any impact on cell cycle movement. MDAMB231 cells were recolored with propidium iodide after 48 h of treatment 50 and 100 µM and exposed to FACS. Histogram of the vehicle (DMSO) rewarded cells demonstrated a standard cell cycle design, which incorporates G1 and G2 isolated by the S stage. The subG1 step (for the most part dead cells) was not noticeable. Endless supply of compound 1, a subordinate focus change was seen in the cell cycle design Figurs 2, 3 and 4).



Fig. 3. Flow Cytometry plots of MDAMB231 cells treated with 50µg/mL of compound Vf



Fig. 4. Flow Cytometry plots of MDAMB231 cells treated with 100µg/mL of compound Vf

Compound Vf has shown S arrest from 19.94% and 32.39%, respectively. G2M phase arrest was found to be 6.38% and 10.44% in MDAMB231 cells. Thus, these outcomes show that compound 1 may meddle with cell division by initiating S stage capture followed by apoptosis. So, more researches are required to comprehend the mechanism of cell cycle capture. that the synthesized 5-benzylidene bis-rhodanine derivatives have a reasonable score and outstanding interactions. The compound (Vc) has an excellent binding score and Compound (Vc), sulphur and the C=O is one of the bisrhodanine moiety form H-bonds with LEU99 and TYR32. It exhibited hydrophobic interactions with GLU100, SER98, ILE15, MET36, GLN12, TYR19, LEU94, LYS68, and GLN95.

Docking Score

From the docking study, it was learned





Fig. 5. 2D and 3D of compound (Vc) with 1DTO. In the 2D structure, the green color circle indicates hydrogen bond interaction

CONCLUSION

In the present work, fourteen new 5-benzylidene bis-rhodanine derivatives (V) and

(Va-m) were have been prepared, characterized by proton NMR, carbon NMR spectroscopy, elemental analysis and evaluated for their anticancer activities against HeLa cell lines by MTT assay, leukemic cell line K562, breast cancer cell line MDAMB231 and molecular docking studies showed better binding scores. Screening of (Vc) and (Vf) compounds was carried for their potential anticancer activity. The results proposed that further study of such compounds with 5-benzylidenebisrhodanine moietymay be interest.

- Whiting, E.; Raje, M. R.; Chauhan, J.; Wilder, P. T.; Eker, D. V.; Hughes, S. J.; Bowen, N. G.; Vickers, G. E. A.; Fenimor, I. C.; Fletcher, S. *Bioorg. Med. Chem. Lett.*, **2018**, *28*, 523-528.
- Mendgen, T.; Steuer, C.; Klein, C. D. J. Med. Chem., 2012, 55, 743-747.
- Hotta, N.; Akanuma, Y.; Kawamori, R.; Matsuoka, K.; Oka, Y.; Shichiri, M.; Toyota, T.; Nakashima, M.; Yoshimura, I.; Sakamoto, N.; Shigeta, Y. *Diabetes Care.*, **2006**, *29*, 1538-1544.
- Nagahara, K.; Anderson, J. D.; Kini, G. D.; Dalley, N. K.; Larson, S. B.; Smee, D. F; Jin, A.; Sharma, B. S.; Jolley, W. B. *J. Med. Chem.*, **1990**, *33*, 407-415.
- Nazaktabar, A.; Lashkenari, M. S.; Araghi, A.; Ghorbani, M.; Golshahi, H. *Int. J. Biol. Macromol.*, **2017**, *103*, 379-384.
- Sortino, M.; Delgado, P.; Juarez, S.; Quiroga, J.; Abonia, R.; Insuasty, B.; Nogueras, M.; Rodero, L.; Garibotto, F. M.; Enriz, R. D.; Zacchino, S. A. *Bioorg. Med. Chem.*, **2006**, *15*, 484-494.
- Kartsev, V.; Shikhaliev, K. S.; Geronikaki, A.; Medvedeva, S. M.; Ledenyova, I. V.; Krysin , M. Y.; Petrou, A.; Ciric, A.; Glamoclija, J.; Sokovic, M. *Eur. J. Med. Chem.*, **2019**, *175*, 201-214.
- Shaikh, M. S.; Kanhed, A. M.; Chandrasekaran,
 B.; Palkar, M. B.; Agrawal, N.; Lherbet, C.;
 Hampannavar, G. A.; Karpoormath, R. *Bioorg. Med. Chem. Lett.*, **2019**, *29*, 2338–2344.
- Chao, L.; Jia-Chun, L.; Ya-Ru, L.; Cheng, G.; Mei-Ling, Z.; Hong-Yan, L.; Xiao-Zhen, L.; Chang-Ji, Z.; Hu-Ri, P. *Bioorg. Med. Chem. Lett.*, **2015**, *25*, 3052-3057.
- 10. Liu, H.; Sun, D.; Du, H.; Zheng, C.; Li, J.; Piao,

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Conflicts of Interest

We declare that we have no conflict of interest.

REFERENCES

- H.; Li, J.; Sun, L. *Eur. J. Med. Chem.*, **2019**, *172*, 163-173.
- Trotsko, N.; Kosikowska, U.; Paneth, A.; Wujec, M.; Malm, A. *Saudi. Pharm. J.*, **2018**, *26*, 568-577.
- 12. Tarahomi , M.; Baharfar, R.; Mohseni, M. *Clin. Microbiol. Infect.*, **2019**, *4*, 1-5.
- 13. Rostam, A. B.; Peyravi, M.; Ghorbani, M. *Appl. Surf. Sci.*, **2018**, *427*, 17-28.
- Kratky, M.; Vinsova, J.; ikova, J. I. S. *Bioorg. Med. Chem.*, **2017**, *25*, 1839–1845.
- Bhatt, H. B.; Sharma, S. Arab. J. Chem., 2013, 10, 1590-1596.
- Abdel Hafez, N. A.; Elsayed, M. A.; El-Shahawi, M. M.; Awad, G. E. A.; Ali, K. A. J. *Heterocycl. Chem.*, **2018**, *55*, 1729-1737.
- Wacothon, K. C.; Ludovic, P.; Anoubilé, B.; Yves-Alain, B.; Rémy, L. G.; Myriam, R.; Anne, C.; Jean, P. B. *Med. Chem. Res.*, **2015**, *24*, 1653-1661.
- Celestina, S. K.; Sundaram, K.; Ravi, S. Bioorg. Chem., 2020, 97, 103640.
- Khan, N.; Gautam, G.; Gupta, A. k. J. D. D. T.,
 2019, *9*, 161-167.
- Andleeb, H.; Tehseen, Y.; Shah, S. J. A,; khan,
 I.; Iqbal, J.; Hameed, S. *RSC Advances.*,
 2016, *6*(81), 77688–77700.
- Sim, M. M.; Ng, S. B.; Buss, A. D.; Crasta, S. C.; Goh, K. L.; Lee, S. K. *Bioorg. Med. Chem. Lett.*, **2002**, *12*, 697-699.
- Talele, T.T.; Arora, P.; Kulkarni, S. S.; Patel, M. R.; Singh, S.; Chudayeu, M.; Kaushik-Basu, N. *Bioorg. Med. Chem.*, **2010**, *18*, 4630-4638.

- Ahn, J. H.; Kim, S. J., Park, W. S., Cho, S.Y., Ha, J. D., Kim, S. S., Kang, S. K., Jeong, D. G., Jung, S. K., Lee, S. H., Kim, H. M., Park, S. K., Lee, K. H., Lee, C. W., Ryu, S. E., & Choi, J. K. *Bioorg. Med. Chem. Lett.*, **2006**, *16*, 2996-2999.
- Ahmed I. Khodaira, A. I.; Awadb, M. K.; Gessonc, J. P; Elshaierd, Y. A. M. M. *Carbohydr. Res.*, **2020**, *487*, 107894.
- Yan, M.; Zhao, J.; Sun, D.; Sun, W.; Wenting, B.
 Z.; Deng, W.; Zhang, D.; Wang, L. *Tetrahedron.* Lett., **2017**, *73*(24), 3355–3362.
- Dago, C. D.; Ambeu, C. N. T.; Coulibaly, W. K.; Bekro, Y. A.; Bekro, J. A. M.; Guevel, R. L.; Corlu, A.; Pierre Bazureau, *J. Chem. Heterocycl. Compd.*, **2017**, *53*(3), 341–349.
- 27. Moorthy B. T.; Ravi S.; Srivastava M.; Chiruvella K.K.; Hemlal H.; Joy O.; Raghavan

S. C. Bioorganic & Medicinal Chemistry Letters., **2010**, *20*, 6297-6301.

- Ravi, S.; Chiruvella, K. K.; Rajesh, K.; Prabhu,
 V.; Raghavan, S. C. *Eur. J. Med. Chem.*, **2010**,
 45, 2748-2752.
- 29. Didwagh, S. S.; Piste, P. B. *J. Chem. Pharm. Res.*, **2013**, *5*(5), 271-274.
- Kamila, S.; Biehl, E. R. *Tetrahedron. Lett.*, 2012, *53*, 3998-4003.
- Braddock, D. C.; Cailleau, T.; Cansell, G.; Hermitage, S. A.; Pouwer, R. H.; Redmond, J. M.; White, A. J. P. *Tetrahedron Asymmetry.*, 2010, *21*, 2911-2919.
- 32. Arafa,W.A.A.; Shaker, R.M.; Rabeh, S.A. *Heterocycles.*, **2016**, *92*, 1224-1243.
- 33. Harshitharaj, P. K.; Kumar, R.; Ravi, S. *J. Chem. Pharm. Sci.*, **2016**, *9*(4), 2478-2482.