

ORIENTAL JOURNAL OF CHEMISTRY

An International Open Access, Peer Reviewed Research Journal

ISSN: 0970-020 X CODEN: OJCHEG 2024, Vol. 40, No.(2): Pg. 401-406

www.orientjchem.org

Extraction with 5-Chlorosalicylaldehyde Thiosemicarbazone of Cu(II) in n-Butyl Acetate and Its Spectrophotometric Study

PRIYANKA MELWIN RODRIGUES^{1*} and SUNETRA RAJENDRA CHAUDHARY¹

¹Department of chemistry, Kishinchand Chellaram College, Mumbai, India. *Corresponding author E-mail: priyankapereira7@gmail.com

http://dx.doi.org/10.13005/ojc/400210

(Received: December 02, 2023; Accepted: March 06, 2024)

ABSTRACT

The highly reactive 5-Chlorosalicylaldehyde thiosemicarbazone (CSTSC) has been employed as an extractive reagent to create a simple and precise spectrophotometric technique for detecting Cu(II) spontaneously. The complex of 5-Chlorosalicylaldehyde thiosemicarbazone and Cu(II) has been quantitatively extracted in n-butyl acetate in the pH range of 5.4–6.4 after equilibrating for 60 second. Up to 5.0 µg mL⁻¹ concentration of Cu(II), the complex of 5-Chlorosalicylaldehyde thiosemicarbazone with Cu(II) in n-butyl acetate follows Beer's law, with its maximum absorbance occurring at 380 nm. By employing the Jobs continuous variation method, the composition 1:1 has been determined for the Cu(II)-CSTSC complex, and this finding is further supported by the mole ratio method. The extracted complex remained stable for 72 hours. 5.48 x 10^{-3} µg cm⁻² was determined to be Sandell's sensitivity, whereas 1.16×10^4 L mol⁻¹ cm⁻¹ was the molar absorptivity. Commercial and synthetic samples have been effectively analysed using this method.

Keywords: Copper, n-butyl acetate, Extraction, CSTSC, Analysis, Pharmaceutical samples.

INTRODUCTION

Copper metal plays an important role in biological systems, apart from its industrial application. Copper is the third most essential element after zinc and iron in the human body. It plays a significant role in haemoglobin synthesis, oxidative phosphorylation, and various biochemical reactions¹. Diseases like low immune function and anaemia can be caused by a shortage of copper, whereas excessive amounts of copper develop gastrointestinal symptoms like abdominal pain, vomiting, and nausea². They also affect aquatic life. Various analytical techniques, like atomic absorption spectrometry³⁻⁶, spectrophotometry⁷⁻⁹, voltammetry¹⁰⁻¹³, ICP-AES¹⁴⁻¹⁶, ICP-MS¹⁷⁻¹⁸ have been used for the determination of copper. In chemistry, solvent extraction is a growingly significant separation method. Due to its ease of usage, quickness, and versatility, it has emerged as one of the most promising techniques for separating metal ions at the trace level. The extractive spectrophotometric study of copper has been reported with several reagents like hydrazone¹⁹⁻²⁰, thiosemicarbazone²¹⁻²⁴, oxime²⁵, imines²⁶⁻²⁸ and thiourea²⁹. Thiosemicarbazones are a class of molecules that possess well-established pharmacological and medicinal qualities, such as antiparasitic, antiviral, antifungal, antitumor,

This is an <a>Open Access article licensed under a Creative Commons license: Attribution 4.0 International (CC- BY). Published by Oriental Scientific Publishing Company © 2018



and antimicrobial actions. Thiosemicarbazone compounds have been extensively used in extraction because of their strong reactivity towards metal ions and extremely stable coloured complexes. The current study focuses on spectrophotometric analysis of copper (II) with 5-chlorosalicylaldehyde thiosemicarbazone as a reagent. For the study of copper, this method offers an extremely reliable and sensitive approach.

MATERIALS AND METHODS

Instruments and Chemicals

A quartz cell of 1.0 cm has been used for absorbance studies on the Jasco V-730 UV-VIS spectrophotometer. pH was adjusted with Equiptronics pH meter. Weighing was done using an electronic Contech balance (CA 64). All chemicals were of AR grade, obtained from commercial suppliers. All of the trials used freshly made, doubly distilled water. Hydrochloric acid and water that was double-distilled were utilised to dissolve Copper (II) chloride and create a 1000 ppm copper solution. Iodometry was used to standardise the solution³⁰. To get a functional solution, portions were diluted every day. The proper amount of 0.2 M NaOH was mixed with 0.2 M KHP to create a buffer solution with a pH of 5.4–6.4.

Solutions of foreign ions

Anions were dissolved in distilled water whereas metal salts were dissolved in appropriate acids, then water was added to a known volume to create stock solutions of various metal ions and anions.

Synthesis of 5-Chlorosalicylaldehyde thiosemicarbazone

Equimolar amounts of each 5-Chlorosalicylaldehyde (1.567 g) in 25 mL of ethanol were condensed with an aqueous ethanolic solution of thiosemicarbazide (0.911 g) by refluxing the reaction mixture in a water bath for five hours. Precipitate was produced by pouring the mixture into crushed ice. After being filtered out and repeatedly cleaned with water, it was recrystallized from ethanol and then dried. 0.1% reagent solution was prepared by using a recrystallized product.

Procedure

Cu(II) solution containing 2.5–50 μ g mL⁻¹ concentration, 1.0 mL of 0.1% (4.35 x 10⁻³ mole/ dm³) reagent in methanol, and phthalate buffer (pH

6.0) 4.0 mL were combined in a 10 mL volumetric flask. After adding distilled water to make the final volume 10 mL, the mixture was transferred into a separating funnel. The equilibration of the mixture was carried out with 10 mL of n-butyl acetate. Both the organic and aqueous layers were separated when equilibrium was achieved. Sodium sulphate in anhydrous form was used to dry the organic phase. Next, using a 1.0 cm-matched cell at a wavelength of 380nm, the absorbance of the solution was determined in comparison to a reagent blank.

RESULT AND DISCUSSION

Absorption spectra

The greenish complex forms after mixing Copper(II) with 5-Chlorosalicylaldehyde thiosemicarbazone. This complex can be quantitatively extracted into n-butyl acetate in the pH range 5.4-6.4. The complex Cu(II)-CSTSC absorption spectra, when measured in n-butyl acetate between 300nm-500nm and at 380nm, exhibit absorption maxima(Fig. 1 Curve A). Because of the reagent's insignificant absorbance at the complex's λ_{max} , the reagent has no impact on the copper determination process (Fig. 1 Curve B). Therefore, for all of the Cu(II)-CSTSC complex absorbance measurements, a 380 nm wavelength was selected.



Fig. 1(A). Cu(II)-CSTSC Complex absorption spectra vs. CSTSC blank (B). CSTSC absorption spectra vs. n-butyl acetate blank

pH Study

The extractions have been performed in n-butyl acetate at different pH to find out the influence of pH on colour development, maximum complexation, and the Cu(II)-CSTSC complex's quantitative extraction. Buffer solutions of pH 1–10 were prepared in series. In the presence of these pH the colour development and subsequent extraction of the Cu(II)-CSTSC complex in n-butyl acetate were carried out as per the above procedure. It was observed that maximum extraction of Cu(II) ions takes place between pH 5.4-6.4 as the organic phase shows maximum absorbance. No absorbance was observed by the diethyldithiocarbamate method in the aqueous phase after decomposition, which indicates the absence of the Cu(II) ion. Therefore, an optimum pH range of 6.0 was maintained for extraction.

Effect of Solvent

Different organic solvents were used to study the influence of solvents on the Cu(II)-CSTSC complex extraction and absorbance. The solvents exhibit varying percentages of copper(II) ion extraction as Cu(II)-CSTSC complex: butyl acetate (99.9%), n-butanol (95.0%), ethyl acetate (85.0%), isoamyl alcohol (80.0%), cyclohexanone (40.0%), toluene (14.9%), benzene (10.00%), chloroform (10.00%), carbon tetrachloride (5.00%), xylene (5.0%), and hexane (2.5%). The Cu(II)-CSTSC complex can be easily extracted in an oxygenated solvent. Since the miscibility of n-butyl acetate with water is less; all extractions were carried out in n-butyl acetate.

Effect of reagent concentration

The reagent concentration required for maximum colour development and quantitative extraction is determined by adding 0.1 mL to 1.5 mL of 0.1% reagent concentration (4.35 x 10^{-3} M) in 30 µg of copper(II) solution. A four-fold excess concentration was required for maximum colour development, and adding excess reagent concentration did not affect the absorbance of the solution. Hence, 1.0 mL of reagent concentration was added to all further studies. The impact of concentration of reagent on Cu(II)-ion extraction is shown in Figure 2.



on Cu(II)-ion extraction



The Cu(II)-CSTSC complex can be extracted completely by equilibrating for 60 second. The extracted complex remained stable for 72 hours. Beer's range and sensitivity Cu(II) at 380 nm between the concentration range that varies from 0.25-5.0 μ g mL⁻¹ obeys Beer's law (Fig. 3). 5.48 x 10⁻³ μ g cm⁻² was determined to be Sandell's sensitivity, and 1.16x10⁴ L mol⁻¹ cm⁻¹ was the molar absorptivity.



Influence of salting out agents on the absorbance

Ammonium sulphate, calcium chloride, magnesium sulphate, sodium chloride, and potassium carbonate were added as salting out agents to increase the extraction of metal complex in n-butyl acetate. It was observed that the absorbance of the solution remains the same after adding a salting out agent.

Composition determination of the Cu(II)-CSTSC complex

By employing the Jobs continuous variation method, the complex composition has been determined, and this finding is further supported by the mole ratio method. Copper(II) and CSTSC solutions were prepared in equimolar concentrations (4.35x10⁻⁴M). The varying proportions of metal and reagents were mixed by keeping the total volume fixed at 2.0 mL. The composition 1:1 of Cu(II)-CSTSC complex was formed, as shown by the plot (Fig. 4) of absorbance at 380 nm against the mole fraction of metal. Fig. 5 illustrates the mole ratio method.



the Complex of Cu(II)-CSTSC



Impact of foreign ions

Cu(II)-CSTSC absorbance was investigated with 30.0 µg of copper(II) and different concentrations for every foreign ion to determine the impact of interfering ions. The absorbance value's tolerance limit was set at an error of $\pm 2\%$.

For the various ions under investigation, the tolerance limits were as follows (Table 1):

 F^- , Cl⁻, Br⁻, l⁻, SO₄²⁻, SO₃²⁻, ClO₃⁻, BrO₃⁻, IO₃, SCN⁻, acetate, tartarate, urea, thiourea (20000µg each), NO₂⁻, NO₃⁻ (10000µg each), S₂O₃²⁻, S₂O₈²⁻ (5000µg each), Ca(II), Mg(II), W(VI), Cr(III), Zn(II) (2000µg each), Al(III), Pb(II), Sn(II), La(III), Sr(II) (1000µg each), U(VI), Th(III), Cd(II), (500µg each), V(V), Ce(IV), Bi(II) (200µg each), and Fe(III) (75µg). Severe interference of Co(II), Hg(II) Pd(II), Ag(I) and EDTA was observed at all concentration levels, and interference of EDTA, Hg(II) Co(II), Pd(II) and Ag(I) can be removed by using a suitable masking agent.

Table 1: Impact of foreign ions on extraction	n of Cu(II)-CSTSC	complex
---	-------------------	---------

Interfering ions	Tolerated amount in μg	Interfering ions	Tolerated amount in µg	Interfering ions	Tolerated amount in µg
F⁻	40000	SCN⁻	10000	Sn ²⁺	1000
Cl⁻	40000	NO_	10000	La ³⁺	1000
Br⁻	40000	NO ^{^-}	10000	Al ³⁺	1000
I-	40000	S,O,2.	5000	U ⁶⁺	500
CIO ₂ -	20000	S_O_2.	5000	Th ³⁺	500
BrO₄⁻	20000	EDTA	Masked	Cd ²⁺	500
IO,-	20000	Ca ²⁺	2000	V ⁵⁺	200
SŐ,2.	20000	Mg ²⁺	2000	Ce4+	200
SO,2-	20000	W ⁶⁺	2000	Bi ²⁺	200
P2022-	20000	Cr ³⁺	2000	Fe ³⁺	75
Acetate	20000	Mn ²⁺	2000	Hg ²⁺	Masked
Tartarate	20000	Zn ²⁺	1000	Co ²⁺	Masked
Urea	20000	Sr ²⁺	1000	Ag ⁺	Masked
Thiourea	20000	Pb ²⁺	1000	Pd ²⁺	Masked

Precision and accuracy

Ten different 30 µg copper containing solutions absorbance were measured in order to determine accuracy and reproducibility. The standard deviation and these ten readings average was computed. It was discovered that the standard deviation was 0.017. The reproducibility of results with a 95% confidence level, based on the standard deviation, was 2.999±0.010.

Applications

Pharmaceutical samples, alloys, and synthetic mixtures are all tested for copper content using the current method.

Determination of copper ion concentration in synthetic mixture

Using the suggested method, copper ions were determined from a number of synthetic mixtures with different copper(II) and other metal ion compositions. Table 2 presents the findings.

Table 2: Copper(II) content analysis in a synthetic mixture

Composition of mixtures in µg	Amount present in µg	Found by present method in µg
Cu(40), Cd(100), Zn(100)	40.0	39.9
Cu(40), Pb(50), Mn(100)	40.0	39.85
Cu(40), Fe(50), Cr(100)	40.0	39.7
Cu(40), Th(100), Zn(50)	40.0	39.8

Detection of Cu(II) ions in alloy materials

The copper content of alloys such as Brass and Devarda's alloys was measured using the proposed technique. 0.1 g of the alloy samples were dissolved in 10 mL of aqua regia after being dried in an oven. Following almost complete drying, 5 mL of dil. HCl was added, and then, up to 100 mL the solution was diluted using double-distilled water. The suggested method was then followed to test the right quantities of these solutions for copper(II). Table 3 displays the findings.

Detection of copper levels in drug samples

10 mL of aqua regia were used for each pharmaceutical sample, and the samples were heated until almost dry. After applying $HClO_4$ to the organic matter to break it down, 10 mL of 2N HNO_3 were used to extract the residue. After filtering, distilled water was added to the solution to dilute it to the desired volume. The suggested method was used to analyse suitable aliquots for copper(II). Table 3 represents the outcomes of analysis.

Table 3: Detection of Cu(II) in drugs and alloys samples (n=4 average of four determination)

Sample Amount present		Amount found	
Keraboost	3.0 mg/tablet	2.98 mg/tablet	
Antoxid-HC	1.0 mg/tablet	0.97 mg/tablet	
Brass alloy	67.9%	67.7%	
Devardas Alloy	48.2%	48.1%	

- 1. Sailer, H.G.; Sigel A.; Sigel, H.; , Marcel Dekker, New York, *Handbook on metals in clinical and analytical chemistry.*, **1994**.
- 2. Scheiber, I; Dringen, R.; Mercer, JF., *Met Ions Life Sci.*, **2013**, *13*, 359-87.
- Hosseini, Mehdi; Dalali, Nasser; Mohammad nejad, Saeid; International Journal of Industrial Chemistry., 2012, 3, 7.
- Yang, Qiang.; Tang, Guang Ping.; Tian, Lun Fu.; Wei, Qi Long.; Wang, Chao.; Advanced Materials Research., 2015, 1120-1121, 1395-1398.
- 5. Han, Quan.; Yang, Xiaohui.; Huo, Yanyan.; Lu, Jiale.; Liu, Yaqi.; *Separations.*, **2023**, *10*(2), 93.
- Soylak, Mustafa.; Unsal, Yunus Emre.; Kizil, Nebiye.; Aydin, Ayse.; *Food and Chemical Toxicology.*, **2010**, *48*, 517-521.
- Babayeva, Könül.; Demir, Serkan.; Andac, Müberra.; *Journal of Taibah University for Science.*, 2017, 11, 808-814.
- Alharthi, Salman. S.; Al-Saidi, Hamed. M.; Appl. Sci., 2020, 10(11), 3895.
- Dorehgiraee, Ali.; Pourabdollah, Elham.; Ghanei-Motlagh, Masoud.; *J. Chil. Chem.* Soc., 2018, 63, 4.
- Squissato, André L.; Richter, Eduardo M.; Munoz, Rodrigo A.A.; *Talanta.*, **2019**, *201*, 433-440.
- 11. Suyanta, Suyanta.; Sunarto, Sunarto.; Padmaningrum, Regina Tutik.; Karlinda,

CONCLUSION

This method provides high sensitivity and selectivity for spectrophotometric detection of Cu(II) in microgram amounts. The extraction was not affected in presence several anions and cations. The amount of Cu(II) can be measured very quickly from alloys and pharmaceutical samples, as the method is simple and rapid.

ACKNOWLEDGMENT

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Conflict of Interest

The authors declare no competing financial interest.

REFERENCES

Karlinda.; Illyas, Md. Isa.; Rahadian, Rahadian.; Indones. J. Chem., **2021**, *21*(2), 332-339.

- 12. Georgieva.; Todorov, Petar, Artem.; Bezfamilnyi.; *Journal of Chemical Technology and Metallurgy.*, **2021**, *56*(5), 999-1007.
- Hassan, Khalid M.; Elhaddad, Ghada M.; Azzem, Magdi Abdel.; *Microchimica Acta.*, 2019, *186*, 440.
- 14. Paktsevanidou, Ioanna P.; Manousi, N.; Zachariadis, G. A.; *Analytical Letters.*, **2021**, *54*(13), 2227-2238.
- 15. Manousi, Natalia.; Zachariadis, George A.; *Separations.*, **2020**, *7*(4), 66.
- Hussain, Adil.; Sajid, Muhammad.; Potter, Daniel.; Rasheed, Hassam.; Hassan, Mujtaba.; Akhtar, Naeem.; Ahmad, Bashir.; Bokhari, Syed Ali Imran.; *Biological Trace Element Research.*, **2023**, *201*, 4143–4155.
- Mohd-Taufek, Nor.; Cartwright, David.; Davies, Mark.; Hewavitharana, Amitha K.; Koorts, Pieter.; Shaw, Paul N.; Sumner, Ronald.; Lee, Eugene.; Whitfield, Karen.; *Food Analytical Methods.*, 2016, *9*, 2068–2075.
- Cao, Yupin.; Feng, Jinsu.; Tang, Lifu.; Yu, Chunhe.; Mo, Guichun.; Deng, Biyang.; *Talanta.*, **2020**, *206*, 120174.
- D., Rekha.; K., Suvardhan.; K., Suresh Kumar.; P., Reddy, Prasad.; B., Jayaraj.; P., Chiranjeevi.; *J. Serb. Chem. Soc.*, **2007**, *72* (3), 299–310.

- 20. Aswar, A. S.; Joshi, M.D.; Indian Journal of Chemical Technology., 2008, 15, 79-81.
- Reddy, B. K.; Kumar, J. R.; Reddy, K. J.; Sarma, L. S.; Reddy, A. V.; *Analytical Sciences.*, **2003**, *19*, 423–428.
- Reddy, K. Janardhan.; Kumar, J. Rajesh.; Narayana, S. Lakshmi.; Ramachandraiah, C.; Thriveni, T.; A. Reddy, Varada.; *Environ Monit* Assess., 2007, 124, 309–320.
- 23. Madan, Parinita Umesh.; Barhate, Vasant Dnyandeo.; *Euro. J. of Biomedical and Pharm. Sciences.*, **2016**, *3*(4), 392-396.
- Reddy, Nagarjuna.; Reddy, Vasudeva.; Momona Ethiopian Journal of Science., 2012, 4(2), 70-83.
- 25. Sreedhar, N.Y.; Jyothi, Y.; Indian Chem. Soc.,

2005, *82*, 1038-1040.

- Shinde, D. R.; Chaskar, M. G.; Tambade, P. S.; International Conference on Frontiers in Life and Earth Science., 2018, 5(1), 38-41.
- 27. Gupta, AnandKumar S.; Barhate, V. D.; *J. Chem. Pharm. Res.*, **2011**, *3*(6), 159-165.
- 28. Patil, Jayashree S.; International Journal of Advanced Research in Science, Communication and Technology., **2022**, *2*, 3.
- Kuchekar, Shashikant R.; Pulatea, Shivaji D.; Aherb, Haribhau R.; Gaikwadc, Vishvas B.; Hand,Sung H.; *Eurasian Chemical Communications.*, **2019**, *1*, 212-222.
- Vogel, A.I.; Longman, Green, London, Vogel's textbook of quantitative chemical analysis., 1961.