

ORIENTAL JOURNAL OF CHEMISTRY

An International Open Access, Peer Reviewed Research Journal

ISSN: 0970-020 X CODEN: OJCHEG 2020, Vol. 36, No.(3): Pg. 513-523

www.orientjchem.org

Variation in Chemical Composition and Antimicrobial Activities of Essential oil of Leaves of Knob Wood, *Zanthoxylum chalybeum* Collected from Three Different Places of Eastern Ethiopia

TADESSE MOLLA TIGNEH* and NEELAIAH BABU G.

Department of Chemistry, College of Natural and Computational Sciences, Haramaya University, Dire Dawa, Ethiopia. *Correponding author E-mail: tadessemt@gmail.com

http://dx.doi.org/10.13005/ojc/360322

(Received: October 14, 2019; Accepted: June 01, 2020)

ABSTRACT

The chemical compositions and functional properties of Zanthoxylum chalvbeum (Knob wood) leave essential oils varied as growing locations. In this study, the variation in chemical composition and antimicrobial activities of the hydro distilled essential oils of the plant fresh leaves collected from three different growing places in Eastern Ethiopia (Awale, Dire Dawa Administration; Awaday, Haramaya subdistrict, and Lange, Kersa Subdistrict, East Hararghe Zone of Oromia Region), were investigated. The chemical composition of the respected essential oils, as determined by the GC-MS analysis, 13, 12 and 17 compounds representing 99.98%, 100.00% and 100.00% of the total leaves essential oil from the plant growing in Awale, Awaday, and Lange, respectively, were identified. The essential oils of Awale, Awaday and Lange were mostly β -phellanderene (23.96%, 45.31%, 23.45% respectively), whereas highest amount of other components in the essential oils of Lange: terpine-4-ol (25.02%) and sabinene (11.73%); Awaday: α-phellanderene (12.85%) and Awale: the sole β-myrcene (29.33%) was identified. In vitro antimicrobial activity in 20 μL dose of each essential oil evaluated by the disc diffusion method indicated strong antibacterial activities against the test bacteria (S.thyphi, Str. agalactia, S.aureus and E.coli) and slight antifungal activities against the test fungi (A.niger, F.oxysporium). The essential oils' major constituents, the monoterpenoids, such as phellandrenes (α and β), terpinen-4-ol and sabinene could be responsible for the significant antimicrobial activities in this study. The plant essential oils' chemical composition and the antimicrobial activities significantly differ with growing locations. The findings confirm the plant's medicinal properties in traditional uses. The results open the many possibilities for further research on biocidal and antimicrobial activities of the essential oil and its individual components of the species growing in different regions of Ethiopia.

Keywords: Zanthoxylum chalybeum, Essential oil, β-Phellandrene, Hydrodistillation, Antimicrobia.

INTRODUCTION

Natural products albeit their chemical composition unknown are utilized by humankind for

thousands of years as sources of various human needs such as medicines, therapeutic needs and drugs¹. Written documents about the knowledge of traditional uses of plants hardly exist² thus obscuring

This is an 👩 Open Access article licensed under a Creative Commons license: Attribution 4.0 International (CC- BY). Published by Oriental Scientific Publishing Company © 2018



the discovery of new drug sources, their sustainable uses 3 and the retention of their actual values by the respective countries of origins. No adequate quantitative data that could be used to assess the indigenous knowledge on the use and management of medicinal plants in Eastern Ethiopia exists⁴. Documenting traditional medicines and remedies is paramount in order to promote their safety, effectiveness, and affordability³.

In Africa, up to 80% of the population uses traditional medicine for primary health care. 25% of modern medicines are made from plants first used traditionally⁵. About 40% of all pharmaceutical presently in use are derived from natural sources (plants, fungi, and other microorganisms, animals, etc.), either used directly or with some modifications⁶. For example, in China, the plant *Artemisia annua* is used to treat resistant malaria; in South Africa, the plant *Sutherlandia microphylla* is used to increase vitality in HIV/AIDS patients³.

Traditional medicines are affordable at a time of soaring health-care costs and nearly universal austerity and help us to cop the relentless rise of chronic non-communicable diseases. The interest in traditional medicine and complementary medicine is growing continuously around the world7. In Ethiopia, 80% of the human population and 90% of livestock rely on traditional medicine8. The widespread use of traditional medicine among the population in Ethiopia could be attributed to cultural acceptability, its attributed efficacy against certain types of diseases, physical accessibility, and economic affordability as compared to modern medicine. Natural products are environmentally friendly alternatives to recurring pathogen-resistant drugs and environmentally hazardous chemicals9. New alternative antibacterial agents for mouth rinses are necessitated as some synthetic mouth rinses suffer, like chlorhexidine, from staining teeth¹⁰ and, like triclosan, from environmentally unfriendly; likewise, chloramphenicol, an effective broadspectrum antibacterial drug, suffers from being a hemotoxic drug¹¹. Traditional medicinal natural products are available and better biodegradable as compared to synthetic agents.¹²

The current research on the *Zanthoxylum* chalybeum species locally known as gadda [in Oromgna language] which is used traditionally to

treat mainly Gofla (swollen body part) in humans and life stocks, is a new kind in Eastern Ethiopia. Gofla is a cancerous disease of chronic origin of infectious (bacterial, viral, fungal, protozoa, parasitic, etc.) and non-infectious diseases. Gofla could be transmitted from mother to child, vector to animals/ human, animals/insects to human; injury incurred to the body part. The plant is also used to treat aphrodisiac problems and the various child ailments (e.g. stomach problems, fever, diarrhea, etc.); as analgesics and antipyretics; in a nutshell as food and medicinal plant. The plant is used to clean respiratory tract airways obstruction, to treat psychiatric, fainting and dizziness problems, to enhance women's fertility, to ease childbirth processes and to maintain oral hygiene. Plant parts (root/root bark/stem bark/leaves/ flowers/seeds) are prepared alone or mixed with other substances (e.g. mixed with food, honey, tea, coffee pericarps, other plants, etc.) in the powder, paste, decoction, juice, and infusion, macerated or smoke form for use. Administration of the plant could be orally (chewing, eating or drinking as tonic drinks), nasally, topically or taking bath with its juices, infusion or aqueous extracts.

In animals, the plant is used to treat various animal ailments including body inflammation, anthrax, diarrhea, Exo and endo parasites, swollen neck, urinary blockage, and mastitis. It is used to enhance reproductive fertility in animals, milk production in cows and honey production in bees. It is used as charcoal and to disinfect milk gourds.

What motivated this research on the plant's leaves was that the Zanthoxylum chalybeum plant parts are being sold in the open market places of Eastern Ethiopia by Market vendors as an important traditional medicinal plant mainly for the treatment of cancerous diseases locally known as Gofla; aphrodisiac problems, severe colds, pneumonia, toothache, etc. The researcher preferred to carry out the research on the leaves of the plant to the roots and stems for the leaves are available throughout the year and to avoid the uprooting of the plant in the course of collecting samples for the research. Since the chemical composition of a given species varies due to intrinsic and extrinsic factors with time, such variations need documentation for the forthcoming various researches and subsequent studies on the medicinally important plants as Zanthoxylum chalybeum species. Therefore, the present study was conducted to identify the chemical composition and the antibacterial potentiality of the plant leaf essential oil.

EXPERIMENTAL

MATERIALS AND METHODS

Collection of the plant material

The plant Zanthoxylum chalybeum was collected in February 2017 from three different places of Eastern Ethiopia namely Awale, Dire Dawa Administration (9.50°N, 42.05°E; 1979 mtr); Awaday, Haramaya Sub-district (9.35°N, 42.03°E; 2009 mtr), Lange, Kersa Sub-district (9.44°N, 41.79°E; 2060 mtr) in East Hararghe Zone, Oromia Region. Awale located in Dire Dawa administrative zone which is an arid region of Ethiopia. The weather is hot for the most of the year. Both Awaday (Haramaya Sub-district, Oromia region) and Lange (Kersa District, Oromia region) located at the edge of plateau. These both locations are cooler as they are located more than 1850 meters from main sea level. Botanical specimens of the plant were identified and deposited at the Herbarium of the School of Plant Science, Haramaya University, Haramaya, Ethiopia. The collected plant was stored at room temperature until further study.13

Methods

Preparation of extracts

The fresh leaves of *Zanthoxylum chalybeum* (200 g) of each sample were packed in an air tight polyethylene bag and were stored in the refrigerator at 4°C.

The essential oil of the fresh leave (200 g of the leaf mixed with 1200 mL distilled water) was extracted by hydro distillation using Clevenger's apparatus at atmospheric pressure for a period of six hours until complete exhaustion¹⁴. The oil was then separated from the aqueous layer with 10 mL of chloroform using a separatory funnel. The chloroform oil mixture was dried with 5 g of anhydrous sodium sulphate (Na₂SO₄) and filtered using Whattman No 1 filter paper and was concentrated by using a sealed amber-colored vial at 4°C in the refrigerator for GC-MS analysis and antimicrobial tests.

Spectrometric analysis of *Zanthoxylum chalybeum* leaves essential oils

The essential oil of Zanthoxylum chalybeum

analysis was done using the GC-MS system, an Agilent Technology Model 7820A fitted with HP-5 MS column (30 m x 0.250 mm internal diameter coated with 5% phenyl and 95% methylpolysiloxane film thickness 0.25 µm, stationary phase. The column temperature was programmed from 50 to 120°C at 20°C/min, 120 to 150°C at 4°C/min, 150 to 250°C at 20°C/min (with 10 min hold time) and 3.5 min solvent delay. The injector and the detector (5977E MSD) temperatures were maintained at 325°C and 350°C respectively. Helium was used as a carrier gas at a flow rate of 1 mL/min. The interface temperature was 280°C. The mass spectrometer was operated at an electron impact of 70 ev with an ion source temperature of 230°C. The constituents of the essential oil were identified by their retention time MH (Mass Hunter) Library search NIST (National Institute of Standard and Technology) 14 Library and by comparison with mass spectra data available in the literature. The percentage of each constituent in the oil was determined based on GC peak areas.

Test for antibacterial activity Inoculum preparation

The test bacteria strains Staphylococcus aureus, Streptococci agalectia, Escherichia coli and Salmonella typhimurium from the stock cultures were streak on Muller Hinton plates and incubated for 24 hours. The bacteria colonies well separated were used as inoculums. The inoculums were transferred with a bacteriological loop to autoclaved Mueller Hinton molten agar cooled to 45°C in a water bath. The contents were mixed by swirling the flasks gently. The medium was then poured to sterile Petri plates, allowed to solidify and used for bio-test¹⁵. For the fungi test, mycelial plug from stock culture was transferred to PDA plates and incubated for 5 to 7 days. Then a spore of fungi was harvested with washing the surface of the colony using 10 mL sterile distilled water and was transferred to 250 mL autoclaved PDA cooled to about 45°C in a water bath. The medium containing spore or mycelia suspension was poured to sterile plates, allowed solidify and was used for the disk diffusion bioassay¹⁵.

Antibacterial and Antifungal activity screening

The essential oil extracts of Zanthoxylum chalybeum leaves from the hydrodistillation were evaluated *In vitro* for antimicrobial activity by using the paper disc diffusion method against two positive bacteria (*Staphylococcus aureus* and *Streptococcus agalectiae*), two *Gram-negative*

bacterium (Escherichia coli and Salmonella typhimurium) and two fungi (Aspergillus niger and Fusarium oxysporum). The bacterium cultures were inoculated into the Muller Hinton Agar (MHA) and incubated at 37°C. Fungal cultures were inoculated into Potato Dextrose Agar (PDA) and incubated at 27°C. The bacteria were obtained from the Ethiopia Public Health Institute, Addis Ababa. The fungi (Aspergillus niger from infected groundnut, and Fusarium oxysporum from infected maize) were obtained in Plant Science Laboratory, Haramaya University, respectively. Chloramphenicol was used as a standard drug against bacteria whereas Tilt was used against fungi. Approximately, 10 mL of sterile MHA (for bacteria) and PDA (for fungi) were poured into sterile culture plates and allowed to set in. From each extract, 20 µL of the oil was loaded to the discs of 6 mm in diameter in three replications. The impregnated paper discs were loaded on the plates. The plates were incubated at 37°C (for bacteria) and 27°C (for fungi) 24 hours. The diameter of the zone of inhibition 9 for each bacteria triplicates and fungus triplicates was measured horizontally and vertically after the incubation period. The net diameter of the zone of inhibition for each oil per each microorganism was recorded. The average measurements were recorded for bioassay analysis. The antifungal and antibacterial activity of the plant extracts were evaluated using the measurement results against the test organisms¹⁵.

RESULTS AND DISCUSSION

Variation in essential oil chemical composition

The components of the Zanthoxylum chalybeum leaves essential oils with their respective percentage composition, molecular formula and retention time as analyzed by GC-MS from three different places namely Awale, Awaday and Lange of Eastern Ethiopia are presented in Table 1, 2 and 3 with their respective chromatograms (Fig. 1, 2 and 3). Thirteen, twelve, and seventeen major components were identified in various proportions from Awale, Awaday, and Lange, respectively accounting for each 100% of the oil composition. The Awale, Awaday, and Lange (99.98%, 100%, and 100%) essential oils constituents are terpenes, 90% of each essential oil constituents are monoterpenes, typical of essential oil composition. Sesquiterpenes, diterpenes, hydrocarbons (and their derivatives) constitute the rest.

The total constituents of the essential oils' terpenoids accounted for *Awale* (98.46%), *Awaday* (90.44%) and *Lange* (83.00%) in differing proportions. Hydrocarbons and their derivatives accounted for *Lange* (17.00%) and *Awaday* (9.56%) far exceeded that of *Awale* (1.52%) showing a contrasting composition variation between them.

The Awale essential oil (Table 1) constitutes 88.28% monoterpenoids, 9.71% of them are oxygenated monoterpenes. The monoterpenoids β -myrcene (29.33%), β -phellandrene (23.96%), terpinen-4-ol (9.71%), sabinene (9.31%), α -phellandrene (7.33%), α -pinene (2.59%), β -ocimene (2.31%), Υ -terpinene (2.26%) and α -terpinene (1.48%); the sesquiterpene hydrocarbon, β -copaen (2.29%); the monocyclic diterpene hydrocarbon camphorene isomers (7.49%) as m-camphorene(4.86%), p-camphorene (3.03%); hydrocarbon derivatives as 3, 3-dimethyl-4-(methylamino)-2-Butanone (1.52%) were identified by the GC-MS analysis. Thus, thirteen components constituting 99.98% of the Awale essential oil composition were identified by the GC-MS analysis.

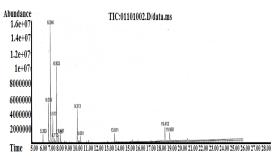


Fig. 1. Chromatogram for essential oil of *Z. chalybeum* leaves, Awale, Dire Dawa

Table 1: Zanthoxylum chalybeum leaves essential oil chemical composition, Awale

			· /	
Peak#	RT	MF	EO Components	%Concentration
1	5.363	C ₁₀ H ₁₆	α -Pinene	2.59
2	6.006	C ₁₀ H ₁₆	Sabinene	9.31
3	6.246	C ₁₀ H ₁₆	β-Myrcene	29.33
4	6.513	C ₁₀ H ₁₆	α -Phellandrene	7.33
5	6.713	C ₁₀ H ₁₆	α -Terpinene	1.48
6	6.933	C ₁₀ H ₁₆	β-Phellandrene	23.96
7	7.188	C ₁₀ H ₁₆	β-Ocimene	2.31
8	7.401	C ₁₀ H ₁₆	Y-Terpinene	2.26
9	9.313	C ₁₀ H ₁₈ O	Terpinen-4-ola	9.71
10	9.651	C ₇ H ₁₅ NO	3,3-dimethyl-4-	
			(methylamino)-2	
			-Butanoned 1.52	
11	13.611	C15H24	β-Copaenb	2.29
12	19.412	C20H22	m-Camphorenec	4.86
13	19.958	C ₂₀ H ₂₂	p-Camphorenec	3.03

'a', 'b' ,'c' ,'d' represents as they are designate

The Awaday plant species leaves essential oil (Table 2) constituted 88.53% monoterpenoids of which 12.89% are oxygenated monoterpenes. In the area, the monoterpenoids β -phellandrene (45.31%), terpinen-4-ol (12.89%), α -phellandrene (12.85%), sabinene (6.36%), Υ -terpinene (3.16%), β -ocimene (2.68%), α -pinene (3.64%) and α -terpinene (1.63%) were identified. Hydrocarbons and their derivatives amounting to 9.56% were identified as cyclodecane (1.95%), 1-octanol (2.06%) and decanal (5.55%). 100.0% of the Awaday essential oil composition was identified using GC-MS analysis.

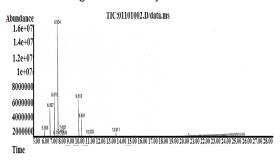


Fig. 2. Chromatogram for essential oil of *Z.chalybeum* leaves, Awaday, Haramaya

 Table 2: Zanthoxylum chalybeum essential oil

 chemical composition, Awaday

Peak#	RT	MF	Components	%Concentration
1	5.365	C10H16	α -Pinene	3.64
2	6.007	C ₁₀ H ₁₆	Sabinene	6.36
3	6.515	C ₁₀ H ₁₆	α -Phellandrene	12.85
4	6.513	C ₁₀ H ₁₆	α -Terpinene	1.63
5	6.934	C ₁₀ H ₁₆	β-Phellandrene	45.31
6	7.189	C ₁₀ H ₁₆	β-Ocimene	2.69
7	7.402	C ₁₀ H ₁₆	Y-Terpinene	3.16
8	7.526	C ₈ H ₁₈ O	1-octanolc	2.06
9	9.313	C ₁₀ H ₁₈ O	Terpinen-4-ola	12.89
10	9.651	C ₁₀ H ₂₀ O	Decanalc	5.55
11	10.603	C ₁₀ H ₁₆	Cyclodecanec	1.95
12	13.611	C ₁₅ H ₂₄	β-Copaenb	1.91

'a', 'b' ,'c' ,'d' represents as they are designated

The Lange Zanthoxylum chalybeum plant species leaves essential oil (Table 3) constitutes 83% monoterpenoids, 25% of them were oxygenated monoterpenes. The major monoterpenoids: 4-terpineol (25.02%), β -phellandrene (23.45%), sabinene (11.73%), α -phellandrene (6.42%), Y-terpinene (5.42%), β -ocimene (5.11%), α -terpinene (2.5%), α -pinene (2.39%), and the minor z-ocimene (0.99%); the hydrocarbons (and their derivatives) amounting 17% as phosphinic amide (2.82%), phenylephrine (2.58%), ethylamine, 2-(adamatan-1yl)-1-methyl- (1.39%), 1-(3,5-dimethyl-1adamatanoyl) semicarbazide (1.67%), benzeneethanamine, 4-methoxy (1.07%), m-methoxyamphetamine (4.98%) were identified. 100.0% of the *Lange* essential oil composition was identified using GC-MS analysis.

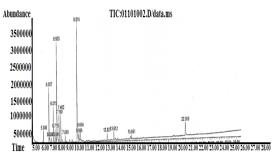


Fig. 3. Chromatogram for essential oil of *Z. chalybeum* Leaves, Lange, Kersa

Table 3: Zanthoxylum	chalybeum	leaves essential
oil chemical o	composition	, Lange

Peak	# RT	MF	Component %	Concentration
1	5.366	C ₁₀ H ₁₆	α-Pinene	2.39
2	6.007	C10H16	Sabinene	9.87
3	6.082	C10H16	Sabinene	1.86
4	6.247	C ₂ H ₄ CINO	Chloroacetamide b	1.27
5	6.515	C ₁₀ H ₁₆	α -Phellandrene	6.42
6	6.715	C ₁₀ H ₁₆	α -Terpinene	2.50
7	6.845	C ₁₀ H ₁₆	z- Ocimene	0.99
8	6.935	C ₁₀ H ₁₆	β –Phellandrene	23.45
9	7.190	C ₁₀ H ₁₆	β-Ocimene	5.11
10	7.402	C ₁₀ H ₁₆	Υ-Terpinene	5.42
11	7.889	C2H4CINO	Chloroacetamide b	1.22
12	9.315	C ₁₀ H ₁₈ O	4-Terpineola	25.02
13	9.505	C2H12NOP	Phosphinic amide b	2.82
14	9.654	$C_9H_{13}NO_2$	Phenylephrine b	2.58
15	12.827	C ₁₃ H ₂₃ N	Ethylamine,	1.39
			2-(adamatan-1-yl)	
			-1-methyl- ^b	
16	13.612	C ₁₄ H ₂₃ N ₃ O ₂	1-(3,5-Dimethyl-1-	1.67
			adamatanoyl)	
			semicarbazideb	
17	15.691	C ₁₀ H ₁₅ NO	Benzeneethanamine, 4-methoxy- ^b	1.07
18	22.09	C ₁₀ H ₁₅ NO	m-Methoxyamphetamine	e [⊳] 4.98
		10 10		

Peak # 4 and 11 are counted as one for they are the same compounds with different RT values. 'a', 'b' represents as they are designated

Chemical composition comparison of the samples from the three places

The chemical composition of the plants collected from *Awale*, *Awaday*, and *Lange* (Table 4) shown insignificant variations with a total composition of monoterpenoid components. Monoterpenoids are the most dominant components in the three places

with almost in the same proportions. But they differ in the composition of individual components. The common monoterpenes percentage of abundance in the order of Awale, Awaday, Lange are α -pinene (2.59%, 3.64%, 2.39%), α-phellandrene (7.33%, 12.85%, 6.42%), and β -phellandrene (23.96%, 45.31%, 23.45%); they are more abundant in Awaday essential oil than Awale and Lange essential oils; sabinene (9.31%, 6.36%, 11.73%), α-terpinene (1.48%, 1.63%, 2.50%), β-ocimene (2.32%, 2.69%, 5.11%), Y-terpinene (2.26%, 3.16%, 5.42%) and terpinen-4-ol (9.71%, 12.89%, 25.02%) are more abundant in Lange than Awale and Awaday essential oils. Solely, β -myrcene (29.33%) appeared in Awale and z-ocimene (0.99%) in Lange area. Sesquiterpene hydrocarbon, β-Copaen, appeared in Awale (2.29%) and Awaday (1.91%) areas. Monocyclic diterpene hydrocarbons, camphorene (7.89%), solely appeared in Awale as m-camphorene (4.86%) and p-camphorene (3.03%). Hydrocarbons and their derivatives appeared: Awale (1.52%) as 3,3-dimethyl-4-(methylamino)-2-Butanone (1.52%); Awaday (9.56%) as cyclodecane (1.95%), decanal (5.55%) and 1-octanol (2.06%); Lange (17.00%) as Ethylamine, 2-(adamatan-1-yl)-1-methyl-(1.39%), 1-(3,5-Dimethyl-1-adamatanoyl) semicarbazide (1.67%) and m-methoxyamphetamine (4.98%) were identified. Thus, the variations confirm the chemical composition of the essential oils from the three places varies as their sources.

The average percentage compositions of the major four common compounds of the respected places are tabulated in Table 5 as shown below.

Referring to Table 5 above, the most abundant terpene of the species in Eastern Ethiopia is β -phellandrene (30.91%), followed by terpinen-4-ol (15.87%), sabinene (9.13%), α -phellandrene (8.87%). β -phellandrene and α -phellandrene are more plentiful in *Awaday* than *Awale* and *Lange*; terpinen-4-ol and sabinene are more plentiful in Lange than *Awale* and *Awaday*. The Eastern Ethiopia *Zanthoxylum chalybeum* is mainly characterized by phellandrene (α and β isomers) on average 40% abundance with β -phellandrene as the main component followed by terpinen-4-ol with 16% and sabinene with 9% average abundance.

Table 4: Zanthoxylum c	halybeum leaves essential
oils chemical co	mposition variation

Identified Compounds	Awale(%)	Awaday(%)	Lange(%)
Monoterpines	_	_	_
α-Pinene	2.59 ^b	3.64ª	2.39°
Sabinene	9.31 ^b	6.36°	11.73ª
α -Phellandrene	7.33 ^b	12.85ª	6.42°
α -Terpinene	1.48°	1.63 ^b	2.50 ^a
β-Phellandrene	23.96 ^b	45.31ª	23.45°
β-Ocimene	2.32°	2.69 ^b	5.11ª
Υ-Terpinene	2.26°	3.16 [♭]	5.42 ^a
β-Myrcene	29.33	_	-
z- Ocimene	-	_	0.99
Terpinen-4-ol	9.71°	12.89 ^b	25.02 ^b
Sesquiterpene			
hydrocarbons			
β-Copaen	2.29 ^a	1.91 ^b	-
Monocyclic Diterpene			
hydrocarbon			
m-Camphorene	4.86	_	-
p-Camphorene	3.03	-	-
Hydrocarbons and			
their derivatives			
3,3-dimethyl-4-(methy	1.52	-	-
lamino)-2-Butanone			
Cyclodecane	-	1.95	-
Decanal	-	5.55	-
1-octanol	-	2.06	-
Ethylamine,2-(adamatan	-	-	1.39
-1-yl)-1-methyl-			
1-(3,5-Dimethyl-1-	-	-	1.67
adamatanoyl) semicarbazide	9		
m-Methoxyamphetamine	-	-	4.98
Benzeneethanamine,	-	-	1.07
4-methoxy-			
Phenylephrine	-	-	2.58
Chloroacetamide	-	-	2.49
Phosphinic amide	-	-	2.82

'-' stands for no compound present under the given place of Zanthoxylum chalybeum. In the designation 'a', 'b', 'c': the values with the same letter are not significantly different (p < 0.05)

Table 5: Representative components of the essential oils for the three places

Compounds	Awale (%)	Awaday (%)	Lange (%)	Averge (%)
1 β-Myrcene	29.33	_	_	_
2 β-Phellandrene	23.96	45.31	23.45	30.91
3 Terpinen-4-ol	9.71	12.89	25.02	15.87
5 Sabinene	9.31	6.36	11.73	9.13
6 α -Phellandrene	7.33	12.85	6.42	8.87
Total	50.31	77.41	66.62	64.78

Though the three areas may have common components but their percentage composition varies with the sources of the essential oil, geographical coordination: *Awale* (latitude: 9.50°N, longitude: 42.05°E, and elevation: 1979 mtr); *Awaday* (latitude: 9.35°N, longitude: 42.03°E, and elevation: 2009 mtr), *Lange* (latitude: 9.44°N, longitude: 41.79°E, and elevation: 2060 mtr). The chemical composition of essential oils is dependent upon the geographical locations for the reasons not known.

As compared to the other African Zanthoxylum chalybeum, the essential oils Zanthoxylum chalybeum of Eastern Ethiopia has a different chemical composition from that of the Eastern and Southern Africa confirming that chemical composition of the species varies as the species sources. The Eastern Ethiopia Zanthoxylum chalybeum leaves oil predominantly on average 71% is monoterpenes as compared to that of Uganda (27%) and Zambia (25%); 16% oxygenated monoterpenes as compare to that of Uganda (61%) and Zambia (66%); with the same total monoterpenoids amounting 88% as compare to Uganda (89%) and Zambia (92%). Zanthoxylum chalybeum leaves oil in Eastern Ethiopia is predominantly consists of monoterpenes while that of Uganda and Zambia predominantly consists of oxygenated monoterpenes. The common major component of Eastern Ethiopia Zanthoxylum chalybeum is β -phellandrene, 31% (average of the three areas) whereas the common major component from the East and Southern Africa (Zambia, Zimbabwe, and Uganda) is citral. Limonene is the major component in Eastern and Southern African countries. It is found that d-limonene induces neoplasms in the kidneys of male rats in association with hyaline droplet nephropathy. Since d-limonene is not in this research findings, the Zanthoxylum chalybeum leaves essential oils under the study is free of neoplasms mentioned earlier in the literature. The Eastern Ethiopia Zanthoxylum chalybeum shares a common terpinen-4-ol with the East and Southern Africa (Zambia, Zimbabwe, and Uganda). The Lange terpinen-4-ol (25.02%) has more or less the same amount as that of Ugandan (22.3%); the Awaleterpinen-4-ol (9.71%) and the Awadayterpinen-4-ol (12.89%) amount twice as that of Zambia (5.0%) and Zimbabwe (5.3%). Thus, the African Zanthoxylum chalybeum differ in composition as their source. The Eastern Ethiopia Zanthoxylum *chalybeum* shares myrcene, α -pinene and sabinene with Zambia; α -pinene, phellandrene (α -, β -) and decanal with that of Uganda. The Eastern Ethiopia Zanthoxylum chalybeum has a different major and common chemical composition from that of the East and Southern Africa Zanthoxylum chalybeum but more or less it approaches to the chemical composition of the Uganda Zanthoxylum chalybeum, especially the Lange Zanthoxylum chalybeum, to some extent and to Zambia to a lesser extent may be attributed to the geological and climatic contrasts of the species sources as well as the differing age of plants.

Antimicrobial Activity of the Essential oils

The antimicrobial activity of the Zanthoxylum chalybeum leaf essential oils extracted by hydrodistillation collected from three different places of Eastern Ethiopia namely Awale, Awaday, and Lange were tested against two Gram-positive (Staphilococcus aureus and Streptococcus agalactiae) and two Gram-negative bacteria (Escherichia coli, Salmonella thyphimurium), chloroform (solvent as negative control) and chloramphenicol (as positive control), as shown in Table 6; as well as two fungi (Aspergillus niger, Fusarium oxysporium) and chloroform (solvent as negative control) and Tilt (Propiconazole, $C_{1r}H_{1r}Cl_{0}N_{0}O_{0}$), as positive control) as shown in Table 7. The disc diameter (6 mm) is excluded for all antimicrobial activities. Chloroform is used as a control to measure its inhibition zone as to subtract it from the total inhibition zone of the solution so as to get the net inhibition zone of the solute, essential oil. In all antimicrobial activities chloroform, the solvent showed no inhibition zone. All inhibition zones recorded were only the net inhibition zones for each oil solute of the plant.

According to Vaquero *et al.*,¹⁶ no antimicrobial activity for inhibition zone<1 mm; antimicrobial activity for inhibition zone \geq 1mm; weakly antimicrobial activity for inhibition zone 1 mm; slight antimicrobial activity, for inhibition zone 2–3 mm; moderate antimicrobial activity, for inhibition zone 4–5 mm; high antimicrobial activity, for inhibition zone 6–9 mm; and strong antimicrobial activity for inhibition zone > 9 mm.

Antibacterial Activities

Referring to Table 6, all test essential oils of the different places and the reference drug chloramphenicol, showed different inhibition zone on each bacterial species. The negative control, chloroform (solvent) did not show inhibition zone on the respective bacteria species. All inhibition values of the test essential oils were entirely for the respected solutes, not the solution.

Essential oil Samples	Dose (µL)	Gram-posi	<i>tive</i> bacteria	Gram-negative bacteria	
		S. aureus	Str.agalactiae	E.coli	S. thyphi
Awale	20	16.50 ± 4.00	17.17 ± 2.89	7.00 ± 0.33a	19.17 ± 1.44
Awaday	20	17.83 ± 2.11	15.00 ± 1.00	5.35 ± 0.87a	21.67 ± 0.89
Lange	20	12.83 ± 4.11	14.35 ± 1.11	6.50 ± 0.67a	16.50 ± 2.00
Chloroform	20	_	-	_	_
Chloramphenicol	20	26.00	29.00	27.00	18.50
% of Inhibition zone		60.46%	53.48%	23.26% a	76.54%

Table 6. Zone of bacterial growth inhibition 9 of the essential oils

'-' stands for no inhibition zone. Values are mean of three replications 9 \pm SD

Zanthoxylum chalybeum species essential oil is a broad spectrum essential oil in that both *Gram-positive* and *Gram-negative* bacteria are found to be strongly sensitive to it. The exception is *E. coli* (*Gram-negative*) bacterium, the least sensitivity of all the bacteria under investigations, showed high sensitive to all tested essential oils. The relatively less sensitivity of the *E. coli* bacterium is due to its impermeable thick murein layer which prevents the entry of inhibitors or else the inherited plasmids of its cell membrane to the essential oils^{17,18}.

Referring to Table 7, *S.thyphi* (*Gram-negative* bacteria, AlZbac = 19.11 mm \pm 1.74) is the most sensitive of all bacteria followed by the two equally sensitive *Gram-positive* bacteria: *S.agalactiae* (AlZbac = 15.51 mm \pm 1.11) and *S.aureus* (AlZbac = 15.72 mm \pm 1.93) to all three essential oil samples. The *E.coli* (*Gram-negative* with AlZbac = 6.28 mm \pm 0.62) bacterium showed the least sensitivity to all essential oil tests. All the values of the inhibition zone of the test essential oils have a value of almost the half value of the reference drug except *S.typhi* showed more or less the same inhibition zone values as the reference drug; and *E.coli* showed far less than the half inhibition zone values of the reference drug. *E.coli* is the least sensitive of all bacteria to

the essential oils. Sensitivities of the essential oils to the *S. thyphi* (*Gram-negative*) and *S.aureus* (*Gram-positive*) bacteria decrease in the order: *Awaday, Awale, Lange.* Sensitivities of the essential oils to the *S.agalactiae* bacteria decrease in the order: *Awale, Awaday*, and *Lange.* Also, sensitivities of the essential oils to the *E.coli* bacteria decrease in the order: *Awale, Lange, Awaday. Lange* essential oils are the least sensitive of all the essential oils tested in all bacteria. The degree of antimicrobial activity of essential oil depends not only on the source of the essential oil but also on the type of bacteria strain under test.

Referring to Table 7, the essential oils of *Awale* (AIZEO = 14.96 mm \pm 3.98) and *Awaday* (AIZEO = 14.96 mm \pm 4.81) have the same sensitivity to all bacteria whereas the essential oils of *Lange* (AIZEO = 12.55 mm \pm 3.02) is the least sensitive of all. *Zanthoxylum chalybeum* leaves essential oil has strong broad spectrum antibacterial properties with AIZEO = 14.16 mm \pm 1.07. A combination of antagonistic, synergistic and additive interactions plays a great role in the antimicrobial activities of the terpene components depending on concentrations and ratios of components.^{19,20}

Table 7: The average inhibition zone (AIZ) for the samples

Essential oils	Dose (µL)	Gram-posi	<i>itive</i> bacteria	Gram-nega	a <i>tive</i> bacteria	Average
of the places		S. aureus	Str.aga	E.coli	S. typhi	(AIZEO)
Awale	20	16.50 ± 4.00	17.17 ± 2.89	7.00 ± 0.33a	19.17 ± 1.44	14.96 ± 3.98
Awaday	20	17.83 ± 2.11	15.00 ± 1.00	5.35 ± 0.87a	21.67 ± 0.89	14.96 ± 4.81
Lange	20	12.83 ± 4.11	14.35 ± 1.11	6.50 ± 0.67a	16.50 ± 2.00	12.55 ± 3.02 ^b
Average(AIZbac)		15.72 ± 1.93	15.51 ± 1.11	6.28 ± 0.62a	19.11 ± 1.74	14.16 ± 1.07

'a' and 'b' indicate significant difference from the others. Mean inhibition zone \pm S.D

Chloramphenicol (0.01 g/4 mL = 50 µg) is used as positive control²¹

Antifungal Activities

All the samples of the *Zanthoxylum* chalybeum leave extracted essential oils were found to be active to all the tested fungi species as shown in Table 8 below. The essential oils from *Awale*, *Awaday* and *Lange* showed anti-*Asperigillus niger* and *Fusarium oxysporium* fungal activity. Awale and *Awaday* showed high whereas *Lange* showed slight anti-*Asperigillus niger* fungal activity. *Awale* essential oil showed slightly whereas *Awaday* and *Lange* essential oils are weakly anti-fusarium oxysporium fungal activity. ¹⁶ *Awale* essential oil relatively better antifungal than *Awaday* and Lange essential oils. The *Zanthoxylum chalybeum* leaves essential oil in Eastern Ethiopia is found to have broad spectrum antifungal activities that vary as the source

 Table 8: Zanthoxylum chalybeum leaves essential oils fungi growth zone inhibition

Sample	Dose (µL)	Asperigillus niger	Fusarium oxsporyum
 Awaday Awale Lange Chloroform Tilt(100 μL/4 mL) 	20 20 20 20 20	6.67 ± 0.52 7.33 ± 1.21 2.50 ± 0.55 a _ 25.33 ± 1.21	$\begin{array}{c} 1.67 \pm 0.52^{a} \\ 3.00 \pm 0.63 \\ 1.00 \pm 0.00^{a} \\ - \\ 26.67 \pm 0.82 \end{array}$

The values represents mean of three replication \pm SD; '-' stands for no inhibition for the chloroform solvent; [Tilt] = 100 μ L/4 mL; 'a' differs from the others

Table 9: AIZ with respect to essential oil and fungus

Sample	Asperigillus niger	Fusarium oxsporyum	AIZEO
1. Awaday	6.67 ± 0.52	1.67 ± 0.52^{a}	4.17 ± 2.09
2. Awale	7.33 ± 1.21	3.00 ± 0.63	5.17 ± 2.58
3. <i>Lange</i>	2.50 ± 0.55^{a}	1.00 ± 0.00^{a}	1.75 ± 0.75
AlZfu	5.50 ± 2.00	1.89 ± 0.74	3.70 ± 1.81

From Table 9, the most active oil against the tested fungi is *Awale* (AIZEO = 5.17 mm \pm 2.58, highly sensitive) followed by *Awaday* (AIZEO = 4.17 mm \pm 2.09, moderately sensitive) and the least active oil is *Lange* (AIZEO = 1.75 mm \pm 0.75, weakly sensitive). *Asperigillus niger* (AIZfu = 5.5 mm \pm 2.00, highly sensitive) shows the most sensitive whereas Fusarium oxsporyum (AIZfu = 1.89 mm \pm 0.74, weakly sensitive) is the least sensitive to the *Z. chalybeum* oils.

The myriad of essential oil bioactivity of the plant species is related to its intrinsic and extrinsic

factors of the plant species. The intrinsic factors are relative percentage composition, lipophilic and hydrophilic properties, structural differences, the stereochemistry of the components and the possible synergistic interactions between them. The extrinsic factors refer to the plant's parts, stage of growth and its environment and the gram type test bacteria against which it is tested. Some compounds might enhance the growth of fungus while inhibiting various bacteria to differing extents.^{22,23,24}

CONCLUSION

All essential oils of Zanthoxylum chalybeum leaves collected from Awale, Awaday and Lange contain similar major common components as α -pinene, sabinene, α -phellandrene, α -terpinene, β -phellandrene, β -ocimene, Υ -terpinene, and terpinen-4-ol terpenoids in various proportions of all, the most prevalent monoterpenes is β -phellandrene and the most prevalent oxygenated monoterpene is terpinen-4-ol in all essential oils of the places.

The essential oil components differ in their component composition as their sources. The most abundant monoterpene found only in *Awale* essential oil is β -myrcene. The minor monoterpene component found in the Lange area essential oil only is z-ocimene. The sesquiterpene hydrocarbons, β -copaen is detected in Awale and Awaday area only as a major component. Monocyclic diterpene hydrocarbon m-camphorene and p-camphorene are found as major components in the Awale area only. Hydrocarbons and their derivatives cyclodecane, decanal and 1-octanol were detected in Awaday essential oil; and 3, 3-dimethyl-4-(methylamino)-2-Butanone was detected in Awale area essential oil only. Most nitrogenous compounds: ethylamine, 2-(adamatan-1-yl)-1-methyl-; 1-(3, 5-dimethyl-1-adamatanoyl) semicarbazide; m-methoxyamphetamine; phenylephrine; benzene ethaneamine, 4-methoxy-; chloroacetamide; phosphinic amide were detected only in Lange essential oil as major components. The three essential oils have the same similarities as they are from the same plant species and differ as to their sources in both the type and chemical composition of their constituents. All components of the essential oil make up for the uses of the physicochemical, antimicrobial and medicinal properties of the species.

Zanthoxylum chalybeum leaves essential oil to possess antimicrobial activity properties²⁵ and has been used for the treatment of various ailments in Ethiopia and abroad as mentioned in most literature the researcher has come across. The current research data obtained directly from the experimental value would confirm satisfactorily that the Ethiopian Zanthoxylum chalybeum leave essential oil is a broad-spectrum antimicrobial and encourage other researchers for further studies.

- Lahlou, M., The Success of Natural Products in Drug Discovery. *Pharmacology & Pharmacy.*, 2013, 4(3), 17-27.
- Getahun, A., Some common medicinal and poisonous plants used in ethiopian folk medicine Addis Abeba University Press: Addis Abeba., 1976, VG 07 & HG 07.
- WHO Legal Status of Traditional Medicine and Complementary/Alternative Medicine: A Worldwide Review., 2001, 200.
- Belayneh, A.; Asfaw, Z.; Demissew, S.; Bussa, N. F., Medicinal plants potential and use by pastoral and agro-pastoral communities in Erer Valley of Babile Wereda, Eastern Ethiopia. *Journal of Ethnobiology and Ethnomedicine.*, **2012**, *8*(1), 42.
- 5. WHO Traditional medicine., **2003**, 4.
- Nkunya M. H. H., H. W., D. H. Bray In Chemical Evaluation of Tanzanian medicinal plants for the active constituents as a basis for the medicinal usefulness of the plants, International Conference on Traditional Medicinal Plants, Arusha, Tanzania, 18-23 February, 1990; Dar Es Salaam University Press: Arusha, Tanzania., **1991**, 11.
- WHO WHO Traditional Medicine Strategy., 2013, 76, 2014-2023.
- Regassa, R., Assessment of indigenous knowledge of medicinal plant practice and mode of service delivery in Hawassa city, southern Ethiopia. *Journal of Medicinal Plants Research.*, 2013, 7(9).
- Carson, C. F.; Hammer, K. A., Chemistry and Bioactivity of Essential Oils. In Lipids and Essential oils as Antimicrobial Agents., 2010, 203-238.
- Van Strydonck, D. A. C.; Slot, D. E.; Van der Velden, U.; Van der Weijden, F., Effect of a chlorhexidine mouthrinse on plaque,

ACKNOWLEDGEMENT

Authors are grateful for the support received from The Department of Chemistry, Haramaya University, Ethiopia and Haramaya University Model School, Haramaya University, Ethiopia.

Conflict of Interest

We declare that there is no conflict of interest

REFERENCES

gingival inflammation and staining in gingivitis patients: a systematic review. *Journal of Clinical Periodontology.*, **2012**, *39*(11), 1042-1055.

- 11. Pongs, O., Chloramphenicol. Springer: Berlin, Heidelberg., **1979**.
- Bedoux, G.; Roig, B.; Thomas, O.; Dupont, V.; Le Bot, B., Occurrence and toxicity of antimicrobial triclosan and by-products in the environment. *Environ Sci Pollut Res Int.*, **2012**, *19*(4), 1044-1065.
- Meniso, B. G.; Boru, A. D.; Ganjinaboyina, n. B., Phytochemical investigation and evaluation of antimicrobial activities of stem bark of morella salicifolia. *Bulletin of the chemical society of ethiopia.*, **2019**, *33*, 293.
- Clevenger, J. F., Apparatus for the determination of volatile oil. *Journal of the American Pharmaceutical Association.*, **1928**, 17(4), 345-349.
- 15. Hutchinson, C. R., Biological methods for studying the biosynthesis of natural products. *Natural Product Reports.*, **1986**, *3*.
- Vaquero, M. J. R.; Alberto, M. R.; de Nadra, M. C. M., Antibacterial effect of phenolic compounds from different wines. *Food Control.*, **2007**, *18*(2), 93-101.
- A, A. I., Conjugal Transferability of Multiple Antibiotic Resistance in Three Genera of Enterobacteriaceae in Nigeria. *Journal of Diarrhoeal Diseases Research.*, **1992**, *10*(2), 93-96.
- Schelz, Z.; Molnar, J.; Hohmann, J., Antimicrobial and antiplasmid activities of essential oils. *Fitoterapia.*, 2006, 77(4), 279-285.
- Cox, S. D.; Mann, C. M.; Markham, J. L., Interactions between components of the essential oil of Melaleuca alternifolia. *Journal of Applied Microbiology.*, 2001, *91*(3), 492-497.

- 20. Vuuren, S. F. v.; Viljoen, A. M., Antimicrobial activity of limonene enantiomers and 1,8cineole alone and in combination. *Flavour and Fragrance Journal.*, **2007**, *22*(6), 540-544.
- Mith, H.; Duré, R.; Delcenserie, V.; Zhiri, A.; Daube, G.; Clinquart, A., Antimicrobial activities of commercial essential oils and their components against food-borne pathogens and food spoilage bacteria. *Food Science & Nutrition.*, **2014**, *2*(4), 403-416.
- 22. Amaral, J. A.; Ekins, A.; Richards, S. R.; Knowles, R., Effect of Selected Monoterpenes on Methane Oxidation, Denitrification, and Aerobic Metabolism by Bacteria in Pure Culture. *Applied and Environmental*

Microbiology., 1998, 64(2), 520-525.

- 23. Cowan, M. M., *Plant Products as Antimicrobial Agents. Clinical Microbiology Reviews.*, **1999**, *12*(4), 564-582.
- Fatoki, O. A.; Onifade, D. A., Use of Plant Antimicrobials for Food Preservation. International Journal of Biological, Biomolecular, Agricultural, Food and Biotechnological Engineering., 2013, 7(12), 1110-1113.
- 25. Ocheng, F. Ugandan medicinal plants used traditionally for oral care: investigation of extracts for anti-bacterial, cytotoxic and anti-inflammatory effects. Makrere University, Kampala, Uganda., **2015**.