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Brief communication

Antimicrobial and Anti-inflammatory properties of Juglans regia Leaves

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ABSTRACT

Antibacterial and Anti-inflammatory activities of methanolic extract of *Juglans regia* L. leaves (Walnut) grown in Libya were investiagted. Hot extraction method by methanol using Soxhlet apparatus was utilized. The antimicrobial effect was studied using agar diffusion method. Human red blood cell membrane stabilization (HRBC) method was used to estimate the anti-inflammatory properties of the leave extract. The extract of leaves exhibited antimicrobial activity against *Staphylococcus aureus*, and *Candida albicans* with a minimum inhibitory concentration (MIC) of 25 and 50 mg/ml, respectively. In addition, the extract concentrations of 20, 40, and 100 µg/mL inhibited lysis of HRBC with values of 25.8%, 35.5%, and 70%, respectively. The results showed that the methanolic extract of *Juglans regia* leaves exhibited antibacterial activity against *S. aureus*, and antifungal activity against *C. albican*, and produced anti-inflammatory activity.

Keywords: Juglans regia leaves, Methanolic extract, Antimicrobial activity, Anti-inflammatory activity

INTRODUCTION

Herbal medicine was and still one of the leading contributions in the treatment of different disease. Many plants were considered as important element in traditional medicine. Last decades, the scientific attention increased to investigate plants for their active components^{1,2}. An example of one of the valuable and medicinally useful herbs is *Juglans regia* L, which belonged to Juglandaceae family. It's usually known as walnut, which is a large tree distributed throughout the world. The *J. regia* root, stem bark, leaves, seeds and cotyledons are used to treat a variety of health problems. Bactericidal properties of the *Juglans* were reported previously for different part of the plant^{3,4,5}. The bark, leaf, seeds and green husks of *Juglans regia* are used as natural treatments in folk medicine. The bark and

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green husks have been analyzed for its antimicrobial, antifungal and anthelmintic activity; which attributed to diverse chemical constituents^{1,6,7}. The plant leaves from Iranian regions were characterized for different activities8. The plant extract of leaves showed significant hypoglycemic action in both short and long term using leave extract⁹. Also, it was found that leaves showed good oral antimicrobial activity in patient suffering from dental carries⁴. A wide range of antibacterial medicines is used to treat these infections. But with the rise of multidrug resistance and the adverse effect accompanied some antibiotics it is preferable to have alternative therapy of herbal origin to minimize the multidrug resistance⁴. The endogensis and exogenesis factors, effect the plant chemical composition. Studies have reported that chemical compounds found in J.regia are different in different climate. The leaves of this plant rich with flavonoids, phenolic acids, essential fatty acids, ascorbic acid, caffeic acid, paracomaric acid and tannins, in addition to important flavonoids as quercetin galactoside and quercetin pantocid derivatives, quercetin arabinoside, quercetin xyloside, and quercetin rhamnoside^{10,11}. In the current study, the methanolic extract of Libyan cultivated J.regia leaves was conducted and investigated for their antimicrobial and anti-inflammatory activities.

MATERIALS AND METHODS

Collection of the leaves

The plant sample of green leaves of Walnut (*Juglans regia* L.) was collected from local area in Tripoli in summer 2017 and authenticated by Taxonomy center of botany division and referenced (voucher specimen number 5971965). The green leaves were air dried in shade at room temperature to produce powder after grinding.

Preparation of leaves extract

A Soxhlet extraction method was performed according to the standard method described in literature. The powdered dried green leaves of the plant (50 g) placed inside a paper timber where the plant material was macerated at room temperature overnight in 250 ml of methanol. Then the plant material was submitted to Soxhlet extraction for 8 h at a maximum temperature of 60°C. The crude extracts were evaporated to obtain the dry extract and the extractive value was calculated. Extractive value (24.4%) determines the amount of active constituents in a given amount of medicinal plant material when extracted with a solvent.

Antimicrobial activity Microorganisms

The antimicrobial effect of methanolic extract obtained from Walnut was tested against four *Gram-negative* bacteria (*Escherichia coli* (ATCC 25922), *Pseudomonas aeruginosa* (ATCC 27853), *Salmonella enterica* (ATCC 25566), and *Klebsiella pneumoniae* (ATCC13883)) and two *Gram-positive* bacteria (*Staphylococcus aureus* (ATCC 25923), *Bacillus subtilis*) and *Candida albicans* fungi. The bacteria were initially reactivated from stock cultures, and maintained on Mueller Hinton broth (MHB) at 37°C for 18 hours.

Agar diffusion method

The antimicrobial properties of methanolic extract used in this study were determined according to the Clinical and Laboratory Standards Institute protocol utilizing agar well diffusion method with some modifications¹². The bacteria were cultured on Mueller-Hinton Agar (MHA) at 37°C for 18 hours. MHA agar plates were inoculated with tested microrganisms at 0.5 McFarland scale, and wells were filled with 50 µl of Walnut methanolic extract in concentration ranging from 100 to 12.5 mg/mL and incubated for 24 h at 37°C, then diameter of inhibition zone was measured. Ciprofloxacin and Fluconazole (1 μ g/mL) were utilized as positive control. The experiments were performed in triplicate and the zone of inhibition was recorded.

Anti-inflammatory activity

The *in vitro* anti-inflammatory activity of methanolic extract was screened as described previously⁵. Mixing freshly drawn human blood in an equal volume of sterilized Alsever's solution, followed by centrifugation at 3000 rpm for 10 min, and reconstituted using normal saline at 10% (v/v) was a first step. Then 1 mL of tested samples were prepared in normal saline at different concentrations (20, 40, 60, 80, 100) µg/mL mixed with 1 mL of 0.2 M phosphate buffer, 1 mL hypo saline, and 0.5 mL of 10% blood suspension, and subsequently incubated at 37°C for 30 min, then centrifuged for 30 min at 3000 rpm. The haemoglobin content in the supernatant

was determined using spectrophotometry at 560 nm. Distilled water and diclofenac sodium were utiliized as control and standard, respectivelly, in which control representing no stability or 100% lysis, the percentage of inhibition of HRBC haemolysis determined by formula.

% Inhibition of Haemolysis = (O. D. of Control- O. D. of Test sample/O. D. of Control) X 100

Where O. D. represents the optical density or absorbance.

RESULTS AND DISCUSSION

Antimicrobial activity

J.regia exhibited many biological activities

owing to their secondary metabolites such as phenolic and flavonoids, however, thier effect are variabe due to the influence of environmental factors^{5,13} and geographical region. Previous studies reported that the Walnut plant has antimicrobial activities against different microorganisms^{4,7}.

In the current study, the leave extract showed antimicrobial activity against *Staphylococcus aureus*, and *Candida albicans* where as; there was no effect on other microbial species (Fig. 1). Generally the extract exhibited a stronger antimicrobial activity and minimal inhibitory concentration (MIC) against *Staphylococcus aureus* (25mg/mL) than *Candida albicans* (50 mg/mL) (Table1).



Fig. 1. Antimicrobial activity of Walnut Methanolic extract. (A) agar diffusion method was performed to examine antimicrobial activity of 100mg/mL (G) and 1mg/mL (H) of Wallnut methanolic extract against *Escherichia coli*, *Klebsiella*, *Salmonella typhi*, *Candida albicans*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and antimicrobial agents (Ciprofloxacin and Fluconazole) were used as positive control with the concentration of 1µg/ lmL. (B) Zone of inhibition measurement

Table	1:	Minimum	inhibition	concentration	of Walnut	
methanolic extract						

	Concentrations of methanolic extract			
Microorganism	100 mg/mL	50 mg/mL	25 mg/mL	12.5mg/mL
Staphylococcus aureus	18.33 mm	14.33 mm	10 mm	-
Candida albicans	15 mm	10.33 mm	-	-

The above results are consistent with some of previous studies in which the leave extract showed antimicrobial properties against *Gram-positive* not *Gram-negative* bacteria as they were resistant to leaves extract. In contrast, previous study found that *Candida albicans* was resistant to *Juglans regia* leaves extract⁷. Moreover, another study reported that alkaloid of methanolic fraction of extract of *Juglans regia* produced moderated inhibition of *Candida albicans* growth⁵.

Anti-inflammatory activity

As presented in Table 2, the results demonstrated protection of the cell membrane in concentration-dependent pattern. The levels of protection of of Walnut methanolic extract observed were 25.8%, 35.5%, and 70% for 20, 40, and 100 μ g/mL concentrations, respectively (Fig. 1). While the membrane stabilization of diclofenac sodium, were 36.6%, 42.6%, and 85% at concentrations of 20, 40, and 100 μ g/mL, respectively.

Table 2: Anti-inflammatory activity of	Walnut				
methanolic extract					

Concentration µg/mL	%Hemolysis of Methanolic extract(% ± SD)	%Hemolysis of Diclofenac sodium(% ± SD)
20	25.8 ± 0.06	36.6 ± 0.006
40	35.5 ± 0.03	42.6 ± 0.007
60	51.6 ± 0.02	55.4 ± 0.005
80	63 ± 0.01	79 ± 0.037
100	70 ± 0.005	85 ± 0.01

CONCLUSION

In summary, the methanolic extract of *J.regia* leaves was examined for its anti-inflammatory and antimicrobial properties. The extract was active against *Gram-positive* bacteria, and fungus, revealing their antimicrobial properties. In addition, the extract produced anti-inflammatory effect. The findings of this study suggest potential use of *J.regia* leaves extract as antimicrobial

- 1. Firenzuoli, F.; Gori, L. *Evid Based Complement Alternat Med.*, **2007**, *4*, 37-40.
- Shojaii, A.; Dabaghian, F. H.; Goushegir, A.; Fard, M. A. Acta Med Iran., 2011, 49, 637-42.
- Bahmani, M.; Rafieian-Kopaei, M.; Hassanzadazar, H.; Saki, K.; Karamati, S. A.; Delfan, B. Asian Pacific journal of tropical medicine., 2014, 7, S29-S33.
- 4. Chopra, R.; Nayar, S.; Chopra, I. New Delhi., **1986**, 2-79.
- 5. Chopra, R. N.; Chopra, I., *Indigenous drugs* of *India*.: Academic publishers., **1933.**
- Ahmad, I.; Beg, A. Z. Journal of ethnopharmacology., 2001, 74, 113-123.
- Noumi, E.; Snoussi, M.; Hajlaoui, H.; Valentin, E.; Bakhrouf, A. European Journal of Clinical Microbiology & Infectious Diseases., 2010, 29, 81.
- 8. Chaleshtori, R. S.; Chaleshtori, F. S.; Rafieian, M. *Turkish Journal of Biology.*, **2011**, *35*, 635-639.
- 9. Kamyab, H.; Hejrati, S.; Khanavi, M.; Malihi,

and anti-inflammatory agent.

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Conflict of interest

The authors have no conflict of interest.

REFERENCES

F.; Mohammadirad, A.; Baeeri, M.; Esmaily, H.; Abdollahi, M. *Cent. Eur. J. Biol.*, **2010**, *5*, 304–309.

- Mohammadi, J.; Mirzaie, A.; Azizi, A.; Roozbehi, A.; Delaviz, H. *Iran S Med J.*, **2012**, *4*, 293–301.
- Amaral, J.S.; Seabra R.M.; Andrade P. B.; Valentão P.; Pereira J. A.; Ferreres F. Food Chemistry., 2004, 88, 373–379.
- Institute, C. a. L. S., Performance Standards for Antimicrobial Susceptibility Testing. Seventh Informational Supplement CLSI document M100-S17., Clinical and Laboratory Standards Institute: 940 West Valley Road, Suite 1400, Wayne, Pennsylvania, USA., 2007.
- Deshpande, R. R.; Kale, A. A.; Ruikar, A. D.; Panvalkar, P. S.; Kulkarni, A. A.; Deshpande, N. R.; Salvekar, *J. P. International Journal of Pharmacy and Pharmaceutical Sciences.*, **2011**, *3*, 200-201.

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