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Study of Isolation, Characterisation and Antimicrobial Activity of High Value Bioactive Compounds from Methanolic Extract of Leaves of Tilkor (*Momordica monadelpha*)

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ABSTRACT

In this paper an attempt has been made to highlight the physicochemical study of methnolic extract of leaves of Tilkor carried out by soxhlet extraction process, phytochemical analysis of the extract, separations, isolation of bioactive components through Thin Layer Chromatography (TLC) as well as column chromatography respectively and characterisation of isolated compound by the means of several spectral analysis such as ¹H NMR, ¹³C NMR, IR, U.V. Mass spectroscopy. The methanolic extract of leaves of the plant (in tropical conditions of Mithilanchal, Bihar, India) reveal the presence of phytochemicals like alkaloids, flavanoids, tannins, saponins, cardiac glycosides, steroids, terpenoids etc. The secondary metaboilities showed antimicrobial activity. The two isolated compound A as - stigmosterol and compound B as tritriaconatane and is also found to have antimicrobial activity.

Keywords: Momordica monadelpha, Tilkor, Physicochemical analysis, Phytochemical analysis, isolation, Characterisation, Stigmosterol, Tritriacontane, Antimicrobial study etc.

INTRODUCTION

Dependence on plants for the essential such as food, clothes & shelter has been of paramount importance in man's life since the human race began. And a time came when people learnt to use plants to cure diseases and relieve physical suffering. After that some plants were appreciated for the herbal treatment as well as major source of new medicine¹. These are called medicinal plants. Further various investigation have been carried out to identify and characterise the high value bioactive components present in medicinal plants²⁻⁷. Monoradica *monadelpha* (Tilkor) is well known plant of Mithilanchal, Bihar (India) having high nutritive and medicinal value . In Mithila it is used for making several dishes. It grows in all parts of India, Tropical Australia, Fiji and throughout the oriental countries. All parts of this plant posses specific medicinal importance and therefore it must contain various high value bioactive components. Its leaves is considered to have antidiabetic character and its paste used in treatment of skin diseases, osteoarthritis and joint paints. Fruit is used for the treatment of liver problem and are anti anaphylactic. Flowers are good anti oxidant, stems are

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anti-hyperglycemic agent and roots are used as health tonic.

So Its chemical standarisation seems essential to identify the chemical constituents. The present Investigation was therefore taken up for the physico-chemical study of the methenolic extract of the leaves of this region, their phytochemical analysis, isolation of the components present using T. L. C. & column chromatography and their characterization using various spectroscopic method and study of antimicrobial activities.

MATERIALS & METHODS

Chemicals and instruments

All Analytical grade solvents and chemicals were used without any purification (Methanol, Silica gel, Calcium sulphate, n-hexane, chloroform, ethyl acetate, petroleum ether, acetone, ethanol, Trimethyl silliane, Argon, D_2O , Liquid Na, Chromium (III), acetyl acetone). A soxhelt extractor is used for the extraction of plant material and separation of its components as well as their isolation were carried out by thin layer chromatography and column chromatography respectively. IR, UV, Mass, ¹H NMR and ¹³C NMR Spectrometer were used for the characterization of chemical constitutents.

Plant material

The leaves of plant momoradica monadelpha [Tilkor] were collected from medicinal plant garden of Shri Himanshu Shekhar Mallik (at Jale) 40 km away from district Darbhanga, Bihar, India.

The leaves are authentified by the experts i Professor Shashi Shekhar Narayan Sinha [International Scientist, Radiation Genetics, Eminent Botanist and Ex. H. O. D. Botany BRA Bihar University,] ii) Professor (Dr.) Sunil Kumar [Principal, Mahendra. Ayurveda College Tulsipur, Daug, Nepal.]

Preparation of plant extract (Soxhlet extraction)

Fresh leaves of Tilkor Momoradica monadelpha] were washed with distilled water. Then it was fully air dried and after it shade dried at room temperature. The dried leaves was then cut and grinded till it get powdered finely. Now the powdered leaves were then subjected to Soxhlet extractor [914/7] with methanol for continuous hot extraction to get the methanolic extract of the leaves.

Determination of Physico-chemical Parameter

Determination of Physico chemical Parameter such as water and alcohol soluble extractive value, total ash content, acid insoluble ash content, moisture content etc. were determined as per guideline given by WHO.⁸

Phytochemical Screening

Preliminary Qualitative and quantative Phyto chemical Screening for the presence of various phytochemicals such as alkaloid, glycoside, phenol flavanoid, saponins, tannins, reducing sugar etc. was carried out by the separated protocol⁹⁻¹¹.

Separation and Isolation

Separation of components from the obtained extract of plant materials were done by "Thin layer Chromatography" (T. L. C.) and isolation of components was done by column chromotography. T. L. C. was performed on a glass plate of silica gel. This layer of absorbent used as stationary phase and solvent used is known as mobile phase.

Characterization

The isolated components have been characterised by several spectral analysis viz – UV, IR, Mass, ¹H NMR and ¹³C NMR spectroscopy.

Antimicrobial Assay

Antimicrobial activity in Methanolic extract of plant sample was determined by agar well diffusion method (NCCL B, 1995). For the growth of bacterial strain natural agar was used while potato dextrose agar was used for the growth of fungi. In the process, plant extract disolved in DMSO at concentration of 15, 30, 60, 120 mg/ml.

The reference antibiotic 25 mg/ml concentrated solution of cephaximin were prepared for each bacterial & fungal strain.

EXPERIMENTAL

Physicochemical Parameter

Calibrated digital pH meter was used to determine the pH of 5% and 20% methanolic extract

through standard method¹⁴⁻¹⁵. Rest all the parameters are being calculated by standard method.

Phytochemical screening

Phytochemical screening of the plant sample was done by the means of standard experimental tests¹².

Extraction

The methanolic extract of fine by powdered dried leaf of Tilkor was prepared by soxhlet extractor using methanol as solvent (by standard method).

Separation & Isolation: Thin Layer Chromatography (TLC) and Column Chromatography

Methnolic plant extract was taken in a beaker (250 mL) and stirred well for 5 hours. Then the solution was filtered and evaporated using Rotory Evaporator. The residue was dissolved in 10 mL of methanol and the extract(10 L) was spotted on TLC plate and the colour of spots were recorded. Silica gel – GF 024 391 was used as absorbent. T.L. C. fingerprint profile was developed by using methanol. The column was then eluted successfully with n – hexane and chloroform respectively through column chromatography and hence component were isolated.

Characterisation

Characterisation of isolated compound was done by using following spectroscopic techniques U.V. Spectroscopy of sample was done by integrating an optical microscope with U.V. optics, monochromator, white light sources and a sensitive detector.

I R spectrum of sample was recorded by passing a beam of infrared light. The amount of light absorbed at each frequency or wave length was measured by the examination of the transmitted light.

The mass fragmentation of the sample was examined by a mass analyzer and detector. The value of indicator quantity was measured by detector and thus provides the necessary data for the calculation of each quantity present.

 $^1\mathrm{H}\,\mathrm{NMR}$ spectra of sample was recorded in methanol solution and $\mathrm{D_2O}$ solvent. TMS was used as reference and chemical shift value for different H – atom was determined.

1 ml plant sample was taken in longer sample tubes (10 nm long in diameter) under high field magnets. Chromium (III) acetyl acetone was taken as relaxation agent and ¹³C NMR spectrum of sample was recorded.

Anitmicrobial Assay

The pure culture of pathogenic bacteria & fungi were obtained from department of Microbiology, Darbhanga Medical College & Hospital.

Viz. aggregate bactor *actinomycet emcouitians* ATCC (12745), *Staphylococcus aureas* ATCC(10835), *Prevotella intermedia* [ATCC (225)], *Shigella shigella* [ATCC (94295)] and *Porphyromonas giugiralis* ATCC [33658] organism were tested on slant of medium containing 3 mg of nutrient agar/150 ml. The slant were incubated at temp. 45°C for 37 h and were stored at 5°C. The inoculum adjusted at 500 m leading to transmission equivalent to 1 x 10 cell /m. The plant dissolved in DMSO and reference antibiotic cephaxium was prepared. Each plate was incubated with 20 g/ml microbial suspension having concentration of 1 x 10⁸ cells. The organisms were tested.

RESULTS & DISCUSSION

The Physico-chemical analysis's of sample of leaves of momoradica monadelpha is given in Table1.

Table 1: Results of various physicochemical parameters of sample of leaves of plant

Parameter	Result
pH of 5% solution of	7.27
Methanolic plant extract	
pH of 20% solution of	6.95
Methanolic plant extract	
Moisture content	1.23%
Total ash	11.71% of dry wt.
Acid insoluble ash	1.05% of total ash
Water soluble ash	4.95% of total ash
Water soluble extractive	6.70% of dry wt.
Alcohol soluble extractive	7.5% of dry wt.

Phytochemical screening

Phytochemical screening of methanolic extract of leaf of the plant is given in Table 2.

 Table 2 : Results of various phytochemical parameters of sample of leaves of plant

Phytochemicals	Test's Name	Result
Alkaloid	Wagner's Test +	
Saponins	Foam Test	
Tannin	Lead acetate Test	++
Steroids	Liebermann-burchard's test	++
Cardiac glycoside	Legal Test	+
Terpenoid	Salkowski test	++
Flavonoids 1. Shinoda test		+++
	2. Alkaline Reagent Test	+++

Thin Layer Chromatography (TLC) and Column Chromatography

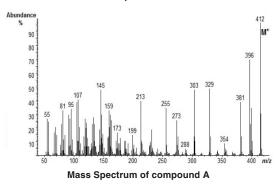
TLC finger print profile was developed by using methanol chloroform & n - hexane solvent in the ratio 0.5:3:4.5 (v/v/v). Six spots were observed (Fig.1) under UV (of 366nm) light when visualized by using vanillin sulphuric acid. Out of the six, two compounds were isolated successfully through the elution with n - hexane and chloroform through column chromatography and were named as compound (A) & compound (B) respectively.



Fig. 1. T.L.C. of sample Spectroscopic Analysis (characterisation) of compound A

Mass Spectroscopy

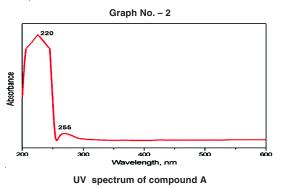
Mass spectrum of compound A showed parent molecular ion [M⁺] peak at m/z 412 which corresponds to molecular formulae $C_{29}H_{48}O$



Graph No. – 1

U.V. Spectroscopy

In U.V. spectral analysis max value of compound A was 255



I R Spectroscopy

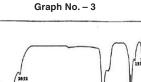
120

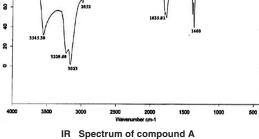
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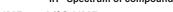
The observed absorption bands of compound A on subjection to IR spectroscopic analysis are given in the Table 3.

Table 3: IR Spectral Data of compound /	Table 3	IR Spectra	I Data of	compound /	A
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S. no.	Observed absorption bands in cm ⁻¹	Inference
1	3545.38 cm ⁻¹	 OH stretching
2	3228.68 cm ⁻¹	Cyclic olefinic – HC = CH –
3	3023 cm ⁻¹	= CH
4	2852.73 cm ⁻¹	C – H
5	1635.81 cm ⁻¹	C = C
6	1460 cm ⁻¹	Cyclic (CH ₂)
7	1378 cm ⁻¹	- CH ₃
8	1070.26 cm ⁻¹	Cycloalkane





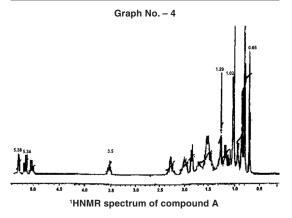


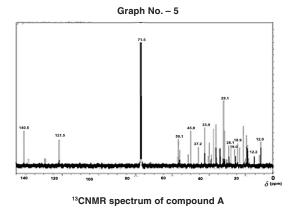


 1 H and 13 C NMR chemical shift value for compound A are recorded in CDCl₃ on the pairs of COSY, HMQC and HNBC correlations [chemical shift values are in (ppm) and computing constants are in Hz] are given in Table 4.

Table 4 : ¹H NMR and ¹³C NMR spectral data of Compound A

$\begin{array}{c ccccccccccccccccccccccccccccccccccc$		Compound A		
$\begin{array}{ccccccc} & & & & & & & & & & & & & & & &$	Position	١H	¹³ C	Nature of carbon
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	C ₁		37.2	CH ₂
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C_2		31.5	CH_2
$\begin{array}{cccccccc} C_{5} & 5.34 \ (+, 1H, J=6.4 \ Hz) & 140.5 & C=C \\ C_{6} & 5.38 \ (S, 1H) & 121.5 & C=CH \\ C_{7} & 32.0 & CH_{2} \\ C_{8} & 32.2 & CH \\ C_{9} & 50.1 & CH \\ C_{10} & 36.5 & C \\ C_{11} & 21.2 & CH_{2} \\ C_{12} & 39.5 & CH_{2} \\ C_{13} & 42.3 & C \\ C_{14} & 56.5 & CH \\ C_{15} & 26.1 & CH_{2} \\ C_{16} & 28.3 & CH_{2} \\ C_{17} & 56.0 & CH \\ C_{18} & 1.29 \ (d,3H) & 36.1 & CH_{3} \\ C_{20} & 33.9 & CH \\ C_{21} & 1.21 \ (d(3H)) & 26.1 & CH_{3} \\ C_{22} & 5.08 \ (m,1H) & 45.9 & C=C \\ C_{23} & 5.21 \ (m,1H) & 23.1 & C=C \\ C_{24} & 0.81 \ (+, 3H \ J=7.1Hz) & 19.0 & CH_{3} \\ C_{26} & 29.1 & CH \\ C_{26} & 0.81 \ (d \ 3H \ = 6.5 \ Hz) & 19.9 & CH_{3} \\ C_{27} & 0.82 \ (d, \ 3H \ J=6.5 \ Hz) & 19.2 & CH_{3} \\ C_{28} & 0.65 \ (S, \ 3H) & 18.9 & CH_{2} \end{array}$	C ₃	3.50(+dd, 1H, J = 4.0, 39Hz)	71.5	СН
$\begin{array}{ccccccc} C_6 & 5.38({\rm S},1{\rm H}) & 121.5 & {\rm C=CH} \\ C_7 & 32.0 & {\rm CH}_2 \\ C_8 & 32.2 & {\rm CH} \\ C_9 & 50.1 & {\rm CH} \\ C_{10} & 36.5 & {\rm C} \\ C_{11} & 21.2 & {\rm CH}_2 \\ C_{12} & 39.5 & {\rm CH}_2 \\ C_{13} & 42.3 & {\rm C} \\ C_{14} & 56.5 & {\rm CH} \\ C_{15} & 26.1 & {\rm CH}_2 \\ C_{16} & 28.3 & {\rm CH}_2 \\ C_{17} & 56.0 & {\rm CH} \\ C_{18} & 1.29({\rm d},3{\rm H}) & 36.1 & {\rm CH}_3 \\ C_{19} & 0.91({\rm d},3{\rm H},{\rm J}=6.1{\rm Hz}) & 19.0 & {\rm CH}_3 \\ C_{21} & 1.21({\rm d}(3{\rm H})) & 26.1 & {\rm CH}_3 \\ C_{22} & 5.08({\rm m},1{\rm H}) & 45.9 & {\rm C=C} \\ C_{23} & 5.21({\rm m},1{\rm H}) & 23.1 & {\rm C=C} \\ C_{24} & 0.81({\rm H},3{\rm H}{\rm J}=7.1{\rm Hz}) & 19.0 & {\rm CH}_3 \\ C_{26} & 0.81({\rm d}3{\rm H}=6.5{\rm Hz}) & 19.9 & {\rm CH}_3 \\ C_{27} & 0.82({\rm d},3{\rm H}{\rm J}=6.5{\rm Hz}) & 19.2 & {\rm CH}_3 \\ C_{28} & 0.65({\rm S},3{\rm H}) & 18.9 & {\rm CH}_2 \end{array}$	C_4		42.3	CH_2
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C_5	5.34 (+, 1H, J=6.4 Hz)	140.5	C=C
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C_6	5.38 (S, 1H)	121.5	C=CH
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C ₇		32.0	CH_2
$\begin{array}{cccccccc} {\rm C}_{10} & & 36.5 & {\rm C} \\ {\rm C}_{11} & & 21.2 & {\rm CH}_2 \\ {\rm C}_{12} & & 39.5 & {\rm CH}_2 \\ {\rm C}_{13} & & 42.3 & {\rm C} \\ {\rm C}_{14} & & 56.5 & {\rm CH} \\ {\rm C}_{15} & & 26.1 & {\rm CH}_2 \\ {\rm C}_{16} & & 28.3 & {\rm CH}_2 \\ {\rm C}_{16} & & 28.3 & {\rm CH}_2 \\ {\rm C}_{17} & & 56.0 & {\rm CH} \\ {\rm C}_{18} & 1.29 ({\rm d},3{\rm H}) & 36.1 & {\rm CH}_3 \\ {\rm C}_{19} & 0.91 ({\rm d}, 3{\rm H}, {\rm J}=6.1{\rm Hz}) & 19.0 & {\rm CH}_3 \\ {\rm C}_{20} & & 33.9 & {\rm CH} \\ {\rm C}_{21} & 1.21 ({\rm d}(3{\rm H})) & 26.1 & {\rm CH}_3 \\ {\rm C}_{22} & 5.08 ({\rm m},1{\rm H}) & 45.9 & {\rm C=C} \\ {\rm C}_{23} & 5.21 ({\rm m},1{\rm H}) & 23.1 & {\rm C=C} \\ {\rm C}_{24} & 0.81 ({\rm +}, 3{\rm H} {\rm J}=7.1{\rm Hz}) & 12.0 & {\rm CH} \\ {\rm C}_{26} & & 29.1 & {\rm CH} \\ {\rm C}_{26} & 0.81 ({\rm d} {\rm 3H}=6.5{\rm Hz}) & 19.9 & {\rm CH}_3 \\ {\rm C}_{27} & 0.82 ({\rm d}, {\rm 3H} {\rm J}=6.5{\rm Hz}) & 19.2 & {\rm CH}_3 \\ {\rm C}_{28} & 0.65 ({\rm S}, {\rm 3H}) & 18.9 & {\rm CH}_2 \end{array}$	C ₈		32.2	CH
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C ₉		50.1	CH
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	C ₁₀		36.5	С
$\begin{array}{ccccc} {\rm C}_{13} & 42.3 & {\rm C} \\ {\rm C}_{14} & 56.5 & {\rm CH} \\ {\rm C}_{15} & 26.1 & {\rm CH}_2 \\ {\rm C}_{16} & 28.3 & {\rm CH}_2 \\ {\rm C}_{17} & 56.0 & {\rm CH} \\ {\rm C}_{18} & 1.29 ({\rm d},{\rm 3H}) & 36.1 & {\rm CH}_3 \\ {\rm C}_{19} & 0.91 ({\rm d}, {\rm 3H}, {\rm J}=6.1 {\rm Hz}) & 19.0 & {\rm CH}_3 \\ {\rm C}_{20} & 33.9 & {\rm CH} \\ {\rm C}_{21} & 1.21 ({\rm d}({\rm 3H})) & 26.1 & {\rm CH}_3 \\ {\rm C}_{22} & 5.08 ({\rm m},1{\rm H}) & 45.9 & {\rm C=C} \\ {\rm C}_{23} & 5.21 ({\rm m},1{\rm H}) & 23.1 & {\rm C=C} \\ {\rm C}_{24} & 0.81 ({\rm +}, {\rm 3H} {\rm J}=7.1 {\rm Hz}) & 12.0 & {\rm CH} \\ {\rm C}_{26} & 0.81 ({\rm d} {\rm 3H}=6.5 {\rm Hz}) & 19.9 & {\rm CH}_3 \\ {\rm C}_{27} & 0.82 ({\rm d}, {\rm 3H} {\rm J}=6.5 {\rm Hz}) & 19.2 & {\rm CH}_3 \\ {\rm C}_{28} & 0.65 ({\rm S}, {\rm 3H}) & 18.9 & {\rm CH}_2 \end{array}$	C ₁₁		21.2	CH_2
$\begin{array}{ccccc} {\rm C}_{13} & 42.3 & {\rm C} \\ {\rm C}_{14} & 56.5 & {\rm CH} \\ {\rm C}_{15} & 26.1 & {\rm CH}_2 \\ {\rm C}_{16} & 28.3 & {\rm CH}_2 \\ {\rm C}_{17} & 56.0 & {\rm CH} \\ {\rm C}_{18} & 1.29 ({\rm d},{\rm 3H}) & 36.1 & {\rm CH}_3 \\ {\rm C}_{19} & 0.91 ({\rm d}, {\rm 3H}, {\rm J}=6.1 {\rm Hz}) & 19.0 & {\rm CH}_3 \\ {\rm C}_{20} & 33.9 & {\rm CH} \\ {\rm C}_{21} & 1.21 ({\rm d}({\rm 3H})) & 26.1 & {\rm CH}_3 \\ {\rm C}_{22} & 5.08 ({\rm m},1{\rm H}) & 45.9 & {\rm C=C} \\ {\rm C}_{23} & 5.21 ({\rm m},1{\rm H}) & 23.1 & {\rm C=C} \\ {\rm C}_{24} & 0.81 ({\rm +}, {\rm 3H} {\rm J}=7.1 {\rm Hz}) & 12.0 & {\rm CH} \\ {\rm C}_{26} & 0.81 ({\rm d} {\rm 3H}=6.5 {\rm Hz}) & 19.9 & {\rm CH}_3 \\ {\rm C}_{27} & 0.82 ({\rm d}, {\rm 3H} {\rm J}=6.5 {\rm Hz}) & 19.2 & {\rm CH}_3 \\ {\rm C}_{28} & 0.65 ({\rm S}, {\rm 3H}) & 18.9 & {\rm CH}_2 \end{array}$	C ₁₂		39.5	CH ₂
$\begin{array}{ccccc} {\rm C}_{14} & 56.5 & {\rm CH} \\ {\rm C}_{15} & 26.1 & {\rm CH}_2 \\ {\rm C}_{16} & 28.3 & {\rm CH}_2 \\ {\rm C}_{17} & 56.0 & {\rm CH} \\ {\rm C}_{18} & 1.29 ({\rm d},3{\rm H}) & 36.1 & {\rm CH}_3 \\ {\rm C}_{19} & 0.91 ({\rm d}, 3{\rm H}, {\rm J}=6.1 {\rm Hz}) & 19.0 & {\rm CH}_3 \\ {\rm C}_{20} & 33.9 & {\rm CH} \\ {\rm C}_{21} & 1.21 ({\rm d}(3{\rm H})) & 26.1 & {\rm CH}_3 \\ {\rm C}_{22} & 5.08 ({\rm m},1{\rm H}) & 23.1 & {\rm C=C} \\ {\rm C}_{23} & 5.21 ({\rm m},1{\rm H}) & 23.1 & {\rm C=C} \\ {\rm C}_{24} & 0.81 ({\rm +}, 3{\rm H} {\rm J}=7.1{\rm Hz}) & 12.0 & {\rm CH} \\ {\rm C}_{26} & 29.1 & {\rm CH} \\ {\rm C}_{26} & 0.81 ({\rm d} {\rm 3H}=6.5 {\rm Hz}) & 19.9 & {\rm CH}_3 \\ {\rm C}_{27} & 0.82 ({\rm d}, {\rm 3H} {\rm J}=6.5 {\rm Hz}) & 19.2 & {\rm CH}_3 \\ {\rm C}_{28} & 0.65 ({\rm S}, {\rm 3H}) & 18.9 & {\rm CH}_2 \end{array}$			42.3	С
$\begin{array}{cccc} {\rm C}_{15} & 26.1 & {\rm CH}_2 \\ {\rm C}_{16} & 28.3 & {\rm CH}_2 \\ {\rm C}_{17} & 56.0 & {\rm CH} \\ {\rm C}_{18} & 1.29 ({\rm d}, {\rm 3H}) & 36.1 & {\rm CH}_3 \\ {\rm C}_{19} & 0.91 ({\rm d}, {\rm 3H}, {\rm J} = 6.1 {\rm Hz}) & 19.0 & {\rm CH}_3 \\ {\rm C}_{20} & 33.9 & {\rm CH} \\ {\rm C}_{21} & 1.21 ({\rm d}({\rm 3H})) & 26.1 & {\rm CH}_3 \\ {\rm C}_{22} & 5.08 ({\rm m}, 1{\rm H}) & 45.9 & {\rm C=C} \\ {\rm C}_{23} & 5.21 ({\rm m}, 1{\rm H}) & 23.1 & {\rm C=C} \\ {\rm C}_{24} & 0.81 ({\rm +}, 3{\rm H} {\rm J} = 7.1{\rm Hz}) & 12.0 & {\rm CH} \\ {\rm C}_{26} & 29.1 & {\rm CH} \\ {\rm C}_{26} & 0.81 ({\rm d} {\rm 3H} = 6.5 {\rm Hz}) & 19.9 & {\rm CH}_3 \\ {\rm C}_{27} & 0.82 ({\rm d}, {\rm 3H} {\rm J} = 6.5 {\rm Hz}) & 19.2 & {\rm CH}_3 \\ {\rm C}_{28} & 0.65 ({\rm S}, {\rm 3H}) & 18.9 & {\rm CH}_2 \end{array}$			56.5	СН
$\begin{array}{cccc} {\rm C}_{16} & 28.3 & {\rm CH}_2 \\ {\rm C}_{17} & 56.0 & {\rm CH} \\ {\rm C}_{18} & 1.29({\rm d},{\rm 3H}) & 36.1 & {\rm CH}_3 \\ {\rm C}_{19} & 0.91({\rm d},{\rm 3H},{\rm J}=6.1{\rm Hz}) & 19.0 & {\rm CH}_3 \\ {\rm C}_{20} & 33.9 & {\rm CH} \\ {\rm C}_{21} & 1.21({\rm d}({\rm 3H})) & 26.1 & {\rm CH}_3 \\ {\rm C}_{22} & 5.08({\rm m},{\rm 1H}) & 45.9 & {\rm C=C} \\ {\rm C}_{23} & 5.21({\rm m},{\rm 1H}) & 23.1 & {\rm C=C} \\ {\rm C}_{24} & 0.81({\rm +},{\rm 3H}{\rm J}=7.1{\rm Hz}) & 12.0 & {\rm CH} \\ {\rm C}_{26} & 29.1 & {\rm CH} \\ {\rm C}_{26} & 0.81({\rm d}{\rm 3H}=6.5{\rm Hz}) & 19.9 & {\rm CH}_3 \\ {\rm C}_{27} & 0.82({\rm d},{\rm 3H}{\rm J}=6.5{\rm Hz}) & 19.2 & {\rm CH}_3 \\ {\rm C}_{28} & 0.65({\rm S},{\rm 3H}) & 18.9 & {\rm CH}_2 \end{array}$			26.1	CH_2
$\begin{array}{cccc} {\rm C}_{18} & 1.29 ({\rm d}, {\rm 3H}) & 36.1 & {\rm CH}_{\rm 3} \\ {\rm C}_{19} & 0.91 ({\rm d}, {\rm 3H}, {\rm J}=6.1 {\rm Hz}) & 19.0 & {\rm CH}_{\rm 3} \\ {\rm C}_{20} & 33.9 & {\rm CH} \\ {\rm C}_{21} & 1.21 ({\rm d}({\rm 3H})) & 26.1 & {\rm CH}_{\rm 3} \\ {\rm C}_{22} & 5.08 ({\rm m}, 1{\rm H}) & 45.9 & {\rm C=C} \\ {\rm C}_{23} & 5.21 ({\rm m}, 1{\rm H}) & 23.1 & {\rm C=C} \\ {\rm C}_{24} & 0.81 (+, {\rm 3H} {\rm J}=7.1{\rm Hz}) & 12.0 & {\rm CH} \\ {\rm C}_{26} & 29.1 & {\rm CH} \\ {\rm C}_{26} & 0.81 ({\rm d} {\rm 3H}=6.5 {\rm Hz}) & 19.9 & {\rm CH}_{\rm 3} \\ {\rm C}_{27} & 0.82 ({\rm d}, {\rm 3H} {\rm J}=6.5 {\rm Hz}) & 19.2 & {\rm CH}_{\rm 3} \\ {\rm C}_{28} & 0.65 ({\rm S}, {\rm 3H}) & 18.9 & {\rm CH}_{\rm 2} \end{array}$	C ₁₆		28.3	CH_2
$\begin{array}{cccc} {\rm C}_{18} & 1.29 ({\rm d}, {\rm 3H}) & 36.1 & {\rm CH}_{\rm 3} \\ {\rm C}_{19} & 0.91 ({\rm d}, {\rm 3H}, {\rm J}=6.1 {\rm Hz}) & 19.0 & {\rm CH}_{\rm 3} \\ {\rm C}_{20} & 33.9 & {\rm CH} \\ {\rm C}_{21} & 1.21 ({\rm d}({\rm 3H})) & 26.1 & {\rm CH}_{\rm 3} \\ {\rm C}_{22} & 5.08 ({\rm m}, 1{\rm H}) & 45.9 & {\rm C=C} \\ {\rm C}_{23} & 5.21 ({\rm m}, 1{\rm H}) & 23.1 & {\rm C=C} \\ {\rm C}_{24} & 0.81 (+, {\rm 3H} {\rm J}=7.1{\rm Hz}) & 12.0 & {\rm CH} \\ {\rm C}_{26} & 29.1 & {\rm CH} \\ {\rm C}_{26} & 0.81 ({\rm d} {\rm 3H}=6.5 {\rm Hz}) & 19.9 & {\rm CH}_{\rm 3} \\ {\rm C}_{27} & 0.82 ({\rm d}, {\rm 3H} {\rm J}=6.5 {\rm Hz}) & 19.2 & {\rm CH}_{\rm 3} \\ {\rm C}_{28} & 0.65 ({\rm S}, {\rm 3H}) & 18.9 & {\rm CH}_{\rm 2} \end{array}$	C ₁₇		56.0	СН
$\begin{array}{cccc} {\rm C}_{_{19}} & 0.91 ({\rm d}, 3{\rm H}, {\rm J}=6.1 {\rm Hz}) & 19.0 & {\rm CH}_{_3} \\ {\rm C}_{_{20}} & 33.9 & {\rm CH} \\ {\rm C}_{_{21}} & 1.21 ({\rm d}(3{\rm H})) & 26.1 & {\rm CH}_{_3} \\ {\rm C}_{_{22}} & 5.08 ({\rm m}, 1{\rm H}) & 45.9 & {\rm C=C} \\ {\rm C}_{_{23}} & 5.21 ({\rm m}, 1{\rm H}) & 23.1 & {\rm C=C} \\ {\rm C}_{_{24}} & 0.81 (+, 3{\rm H} {\rm J}=7.1{\rm Hz}) & 12.0 & {\rm CH} \\ {\rm C}_{_{25}} & 29.1 & {\rm CH} \\ {\rm C}_{_{26}} & 0.81 ({\rm d} 3{\rm H}=6.5 {\rm Hz}) & 19.9 & {\rm CH}_{_3} \\ {\rm C}_{_{27}} & 0.82 ({\rm d}, 3{\rm H} {\rm J}=6.5 {\rm Hz}) & 19.2 & {\rm CH}_{_3} \\ {\rm C}_{_{28}} & 0.65 ({\rm S}, {\rm 3H}) & 18.9 & {\rm CH}_{_2} \end{array}$		1.29 (d,3H)	36.1	CH_3
$\begin{array}{ccccc} C_{20} & & 33.9 & CH \\ C_{21} & 1.21 (d(3H)) & 26.1 & CH_3 \\ C_{22} & 5.08 (m,1H) & 45.9 & C=C \\ C_{23} & 5.21 (m,1H) & 23.1 & C=C \\ C_{24} & 0.81 (+, 3H J=7.1Hz) & 12.0 & CH \\ C_{25} & & 29.1 & CH \\ C_{26} & 0.81 (d 3H=6.5 Hz) & 19.9 & CH_3 \\ C_{27} & 0.82 (d, 3H J=6.5 Hz) & 19.2 & CH_3 \\ C_{28} & 0.65 (S, 3H) & 18.9 & CH_2 \end{array}$		0.91 (d, 3H, J = 6.1 Hz)	19.0	CH₃
$\begin{array}{cccc} {\rm C}_{_{21}} & 1.21 \ ({\rm d}(3{\rm H})) & 26.1 & {\rm CH}_{_3} \\ {\rm C}_{_{22}} & 5.08 \ ({\rm m},1{\rm H}) & 45.9 & {\rm C=C} \\ {\rm C}_{_{23}} & 5.21 \ ({\rm m},1{\rm H}) & 23.1 & {\rm C=C} \\ {\rm C}_{_{24}} & 0.81 \ ({\rm +},3{\rm H}{\rm J}=7.1{\rm Hz}) & 12.0 & {\rm CH} \\ {\rm C}_{_{25}} & 29.1 & {\rm CH} \\ {\rm C}_{_{26}} & 0.81 \ ({\rm d}3{\rm H}=6.5{\rm Hz}) & 19.9 & {\rm CH}_{_3} \\ {\rm C}_{_{27}} & 0.82 \ ({\rm d},3{\rm H}{\rm J}=6.5{\rm Hz}) & 19.2 & {\rm CH}_{_3} \\ {\rm C}_{_{28}} & 0.65 \ ({\rm S},3{\rm H}) & 18.9 & {\rm CH}_{_2} \end{array}$			33.9	СН
$\begin{array}{cccc} C_{22} & 5.08 \ (m,1H) & 45.9 & C=C \\ C_{23} & 5.21 \ (m,1H) & 23.1 & C=C \\ C_{24} & 0.81 \ (+, 3H \ J=7.1Hz) & 12.0 & CH \\ C_{25} & 29.1 & CH \\ C_{26} & 0.81 \ (d \ 3H=6.5 \ Hz) & 19.9 & CH_3 \\ C_{27} & 0.82 \ (d, \ 3H \ J=6.5 \ Hz) & 19.2 & CH_3 \\ C_{28} & 0.65 \ (S, \ 3H) & 18.9 & CH_2 \end{array}$		1.21 (d(3H))	26.1	CH ₃
$\begin{array}{ccc} C_{_{24}} & 0.81 \ (+, \ 3H \ J=7.1 \ Hz) & 12.0 & CH \\ C_{_{25}} & 29.1 & CH \\ C_{_{26}} & 0.81 \ (d \ 3H=6.5 \ Hz) & 19.9 & CH_{_3} \\ C_{_{27}} & 0.82 \ (d, \ 3H \ J=6.5 \ Hz) & 19.2 & CH_{_3} \\ C_{_{28}} & 0.65 \ (S, \ 3H) & 18.9 & CH_{_2} \end{array}$	C ₂₂	5.08 (m,1H)	45.9	C=C
$\begin{array}{ccc} C_{25} & & 29.1 & CH \\ C_{26} & 0.81 \mbox{ (d } 3H = 6.5 \mbox{ Hz}) & 19.9 & CH_3 \\ C_{27} & 0.82 \mbox{ (d, } 3H \mbox{ J} = 6.5 \mbox{ Hz}) & 19.2 & CH_3 \\ C_{28} & 0.65 \mbox{ (S, } 3H) & 18.9 & CH_2 \end{array}$	C ₂₃	5.21 (m,1H)	23.1	C=C
$\begin{array}{ccc} C_{26} & 0.81 \mbox{ (d } 3H = 6.5 \mbox{ Hz}) & 19.9 & CH_3 \\ C_{27} & 0.82 \mbox{ (d } 3H \mbox{ J} = 6.5 \mbox{ Hz}) & 19.2 & CH_3 \\ C_{28} & 0.65 \mbox{ (S, 3H)} & 18.9 & CH_2 \end{array}$	C ₂₄	0.81 (+, 3H J = 7.1Hz)	12.0	СН
$\begin{array}{lll} C_{_{27}} & 0.82 \ (d, 3H \ J = 6.5 \ Hz) & 19.2 & CH_{_3} \\ C_{_{28}} & 0.65 \ (S, 3H) & 18.9 & CH_{_2} \end{array}$	C ₂₅		29.1	CH
C ₂₈ 0.65 (S, 3H) 18.9 CH ₂	C ₂₆	0.81 (d 3H = 6.5 Hz)	19.9	CH_3
		0.82 (d, 3H J = 6.5 Hz)	19.2	-
C ₂₉ 1.02 (S, 3H) 12.2 CH ₃		0.65 (S, 3H)	18.9	
	C ₂₉	1.02 (S, 3H)	12.2	CH3





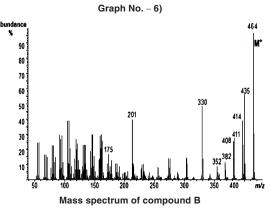
Spectroscopic Analysis (Characterisation) of compound B

UV Spectroscopy

In v. v. spectroscopy the max value of compound (B) is found to be negligible i.e. it showed no absorption.

Mass spectroscopy

Mass spectrum of compound B showed parent molecular ion [M⁺] peak at m/z 464 which corresponds to the molecular formulae $C_{33}H_{68}$. m/z relative intensity are 435 (12,93), 414(11,98) 444(33,72), 408 (22,96), 38(82,03), 352 (100), 330 (53,29), 201(31,33), 175 (18,35) (Graph No. 6).



IR. Spectroscopy

The absorbed absorption band of compound (B) upon subjection to IR spectrometer are given. IR (KBR) ν_{max} (cm⁻¹) : 720 cm⁻¹, 802 cm⁻¹, 865 cm⁻¹, 1025 cm⁻¹, 1095 cm⁻¹, 1260 cm⁻¹, 1375 cm⁻¹, 1465 cm⁻¹, 2845 cm⁻¹, 2915 cm⁻¹ (Graph No. – 7).

¹H NMR Spectroscopy

¹H NMR spectroscopic analysis of compound (B) is as

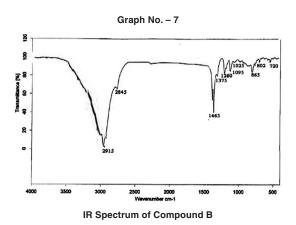
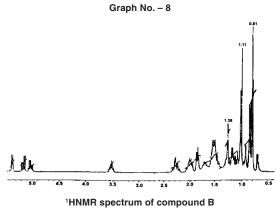


Table 5: ¹H NMR spectral data of compound B

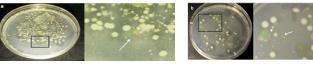
SI. No	o. 1H NMR (δ, CDCl ₃)	Nature of carbon
1	1.11 – 1.38 (62H, br, 31 X)	- CH ₂ -
2	0.81 (6H, + , J 7.4 Hz, 2X)	- CH ₃ -

Pharmacological Activity

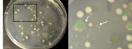


However the % of water soluble ash is greater (4.95) in comparision to acid insoluble ash (1.05%). Among water and alcohol soluble extractives alcohol soluble extractive is greater (75%) than water soluble extractive (6.70%) Phytochemical screening of plant extract carried out with the help of several chemical tests to determine the phytochemicals present in the

SI. No	Organism	Zone of in 15.0	nhibition 30.0	(nm) in 60.0	Conc. mg/ml 120.0	Reference antibiotic 20 μg/ml
1	aggregatibacter actinomycetemcomitans				9.5	14.0
2	Staphylococcus aureus			8.2	10.3	15.0
3	Prevotella intermedia				8.1	10.0
4	Shigella shigella				8.2	11.0
5	Porphyromonas gingivalis			13.2	15.3	13.0



Aggregatibacter actinomycetemcomitans



Staphylococcus aureus



Prevotella intermedia

Porphyromonas gingivalis



Fig. 2. The pictures of incubated palates and micro-organism are given

DISCUSSION

Results obtain from of Physicochemical studies of leave sample of plant to determine the moisture content total ash, acid insoluble ash, water soluble ash water soluble extractives and alcohol soluble extractives along with pH of 5% and 20% solution of methanolic extract of plant leaves were given in Table 1. The pH of 5% solution is greater (7.27) in comparison to pH of 20% solution (6.95). extract, shows the presence of alkaloids saponins tannins, steroid, cardiac glycosides, terpenoids and flavonoids. Out of which alkaloids and flavonoid showed higher degree of precipitation (+++), saponins, tannins, steroid and Terpenoid showed moderate degree of precipitation (++) and cardiac glycoside showed lesser degree of precipitation (+).

Separation and isolation of the plant extract using thin layer chromatography by described method and visualised under U.V. showed six spots out of which two are successfully eluted by n –hexane and chloroform and isolated through column chromatography which are named as compound (A) & (B) and are subjected to several spectroscopic analysis for their characterisation.

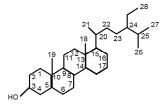
Compound (A) was also isolated as white powder whose mass spectral data. Corresponds to its molecular formulae $C_{2a}H_{50}O$

UV band λ max = 25.5 nm revels the – OH chromophoric group. As it give positive test for steroid, so compound must containing sterol nucleus. The IR absorption bands observed at 35A (5.38 cm⁻¹) that is characteristics of – OH stretching, 3228.68 cm⁻¹ (cyclic olefinic – CH = CH₃) 3023 cm⁻¹ (= CH -), 2852.73 cm⁻¹(C – H), 1635.81 cm⁻¹ (C=C), 1460 cm⁻¹ cyclic (CH₂), 1378 cm⁻¹ (CH₃) and 1070.26 cm⁻¹ (cycloalkane).

The ¹H NMR showed the proton of H - 3 shows multiplet at 3.50 and indicates the existence of signals for olefinic proton at 5.38, 5.08, 5.21 & 5.34. The angular methyl proton at 1.29 & 0.91 correspondence to G8 & C19.

¹³C NMR at 140.5 & 121.5 which are assigned C5 & C6 double bonds respectively. It shows value at 19.0, 33.9 & 26.1. For C19, C20 & C26 corresponds to angular carbon. Four alkenes carbon were appeared at 140.5, 121.5, 45.9 & 23.1 spectra showed twenty nine carbon signal including six methyl, nine methylenes, 11 methanes and three quaternary carbon.

The spectroscopic data corresponds to the structure of compound A as



 β -stigmosterol (compound A)

Compound (B) was isolated as waxy solid mpt(71°C). From IR absorption band data the functional group such as CO & OH are absent. As it showed no absorption in UV spectrum hence the compound is fully saturated.

The mass spectroscopy of compound at 464 corresponds to molecular formula $C_{_{33}}H_{_{68}}$. The ¹H NMR spectral data of compound showed a triplet at 0.81 integrating to 6 proton which may be two (CH₃) at terminals. A multiplate was observed at 1.11 – 1.38 corresponds 62 proton which represent 31 (CH₂) methylene. On the basis of these spectral data the structure of compound (B) is

Tritriacontane Compound (B)

Tritriacontane Compound (B)

Antimicrobial assay of methanolic extract of leaves of the plant exhibit higher antimicrobial activities at 120 mg/ml conc. extract against p. gingiralis (15.3 nm) as compared to reference antibiotic. Antimicrobial activities against other test organism is very less in comparision to reference antibiotic.

CONCLUSION

The methanolic extract of leave of the plant (in tropical conditions of Mithilanchal, Bihar, India) reveal the presence of phytochemicals like alkaloids, flavonoids, tannins, saponins, cardiac glycosides, steroids, terpenoids etc.

The secondary metabolites shows pharmacological activity such as Antimicrobial.

The two isolated compounds were characterised by spectroscopic techniques which revealed the structure of compound A as-stigmosterol and compound B as Tritriacontane

ACKNOWLEDGMENT

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