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# Towards the First Total Synthesis and Anticancer Screening of Polycarponin C: A Cyclic Octapeptide

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### ABSTRACT

The present study describes designing, synthesis and anticancer screening of a proline rich cyclic octapeptide polycarponin C, by solution phase synthesis. The synthesis was carried out by coupling a tetrapeptide Boc-Pro-Thr-Leu-Pro-OH with another tetrapeptide Boc-Pro-Val-Leu-Phe-OH, followed by cyclization of the linear octapeptide. The structure of the synthesized compound was then confirmed by spectral analysis. From the results of biological activity, it was concluded that the compound has shown moderate activity against SR human tumor cell lines of Leukemia when compared with Vincristine as a standard.

**Key words:**Cyclic Peptide, Solution phase synthesis, p-nitro phenyl ester Method, Coupling Agent, Anticancer.

### **INTRODUCTION**

Anticancer drug development from natural sources has always been fascinating and interesting for researchers working in the field of medicinal chemistry and drug development. Owing to the various biological activities like antimicrobial, cytotoxic, anti-HIV, anti-inflammatory, serine protease and protein tyrosine phosphatase inhibitory activity possessed by cyclic peptides<sup>1-7</sup> and to obtain a natural bioactive peptide in good yield, in the present work an attempt has been made towards the first total synthesis of a proline rich cyclic octapeptide polycarponin C, cyclo-( Pro-Thr-Leu-Pro-Pro-Val-Leu-Phe), isolated from whole plants of *Polycarpon prostratum*, belonging to family Caryophyllaceae<sup>8</sup>. The synthesis was attempted by solution phase technique<sup>9</sup> followed by cyclization of linear octapeptide by p-nitro phenyl ester method<sup>12</sup>. The structure of the synthesized compound was

confirmed by detailed spectral analysis. The synthesized compound was then subjected for preliminary cytotoxic activity by using Brine shrimp assay, and further followed by screening against 60 human tumor cell lines at NCI, USA. The results of anticancer screening showed that the compound is moderately active against SR human tumor cell lines of Leukemia when compared with Vincristine as a standard.

### MATERIALS AND METHODS

All L-amino acids, di-*tert*butyldicarbonate (BOC<sub>2</sub>O), diisopropylcarbodiimide (DIPC), trifluoroacetic acid (TFA), triethylamine (TEA), pyridine and *N*-methylmorpholine (NMM) were purchased from Spectrochem Limited (Mumbai, India).Melting points were determined by using digital melting point apparatus.

The IR spectra were run on FTIR spectrophotometer,JASCO 4100 using a thin film supported on KBr pellets or utilizing chloroform and NaCl cells.<sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded on Bruker AC NMR spectrometer using DMSO as a solvent. The mass spectrum of the cyclopeptide was recorded on JMS-DX 303 Mass spectrometer operating at 70 eV by ESIMS/MS.

In order to carry out the total synthesis of cyclicpeptide, polycarponin C, cyclo-(Pro-Thr-Leu-Pro-Pro-Val-Leu-Phe), it was disconnected into four dipeptide units, BOC-Pro-Thr-OMe 1, Boc-Leu-Pro-OMe 2, Boc-Pro-Val-OMe 3, Boc-Leu-Phe-OMe 4. The dipeptides were obtained by coupling Boc amino acids with the respective amino acid methyl esters, by using DIPC as a coupling agent. The ester group

of dipeptide 1 and 3 was then removed by using LiOH and the Boc group of dipeptide 2 and 4 was removed by using TFA. The deprotected units were then coupled to get two tetra peptides Boc-Pro-Thr-Leu-Pro-OMe 5 and Boc-Pro-Val-Leu-Phe-OMe 6. The resulting tetrapeptides were then coupled together by using DIPC and chloroform to obtain a linear octapeptide, which was then cyclized by using p-nitro phenyl ester method to get titled compound.

# General method for preparation of di/tetra/linear octapeptide

L-Amino acid methyl ester hydrochloride/ dipeptide methyl ester/tetra peptide methyl ester (10 mmol) was dissolved in chloroform (CCl<sub>3</sub>, 20 ml). To this, TEA (2.8 ml, 20 mmol) was added at 0 °C and the reaction mixture was stirred for 15 min. Boc-L-amino acid/Bocdipeptide/ Boctetrapeptide (10 mmol) in chloroform (20 ml) and DIPC (10 mmol) were added with stirring. After 24 h, the reaction mixture was filtered and the residue was washed with chloroform (30ml) and added to the filtrate. The filtrate was was hed with 5% NaHCO<sub>3</sub> and saturated NaCl solutions. The organic layer was dried over anhydrous Na SO4, filtered and evaporated in vacuum. The crude product was recrystallized from a mixture of chloroform and petroleum ether (b.p. 40-60 °C) followed by cooling at 0 °C. By using above procedure, compounds 1-7 were synthesized.

# Procedure for cyclization of linear octapeptide<sup>12</sup>:

The cyclization of linear octapeptide was attempted by using p-nitrophenyl ester method. The ester group of linear fragment was removed

Compound	Conc. (ppm or µg/ml)	Num Na	ber of Surv pulii After 2	iving 4 h	Total Number of Survivors	% Mortality	
		T1	T2	Т3			
	1000	4	6	4	14	53.33	
	500	5	5	5	15	50	
Poly C	250	5	6	7	18	40	
	125	7	6	7	20	33.33	
	62.5	8	9	8	25	16.66	
	31.25	10	10	10	30	0	

Table 1: Results for Cytotoxic activity by using Brine Shrimp Assay:

\*T=Trial LC50=460.9 µg/ml

with LiOH and the p-nitrophenyl ester group was introduced. For introduction of p-nitro phenyl ester group, the Boc-peptide carboxylic acid(1.5 mmol) was dissolved in chloroform (15 ml) at 0 °C, to which p-nitrophenyl(0.27 gm, 2 mmol) was added, and stirred for 12 hrs at RT. The reaction mixture was filtered and the filtrate was washed with NaHCO<sub>3</sub> solution (10%) until excess of p-nitrophenyl was removed and finally washed with 5% HCl (5 ml) to get Boc-peptide-pnp ester.

To the above Boc-peptide-pnp-ester (1.2 mmol) in CHCl<sub>2</sub> (15ml), CF<sub>2</sub>COOH (0.274g,

2.4 mmol) was added, stirred for 1 hour at room temperature and washed with 10% NaHCO<sub>3</sub> solution. The organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. To the Boc-deprotected peptide-pnp-ester in CHCl<sub>3</sub> (15ml), N-methyl morpholine (1.4ml, 2mmol.) was added and kept at 0°C for 7 days. The reaction mixture was washed with 10% NaHCO<sub>3</sub> until the byproduct p-nitrophenyl was removed completely and finally washed with 5% HCl (5ml). The organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. Chloroform and pyridine were distilled off to get the crude cyclized compound, which was recrystallized from CHCl<sub>3</sub>/n-hexane.



Graph 1: DTP One dose Mean graph for Polycarponin C

## Anticancer activity of synthesized compound Preliminary cytotoxic activity by Brine Shrimp Lethality Assay (BSLA)<sup>10, 11</sup>

Brine shrimp eggs were obtained from the aquarium shop, Nasik. Artificial sea water was prepared from (1% NaCl) nitrate, phosphate, and silicate-free sea-salt and distilled water (35 g/l) at 25° C under constant illumination. The saltwater solution was aerated continuously during incubation with an aquarium air pump. The seawater was put in a small plastic container (hatching chamber) with a partition for dark (covered) and light areas. Shrimp eggs were added into the dark side of the chamber while the lamp above the other side (light) will attract the hatched shrimp. Two days were allowed for the shrimp to hatch and mature as nauplii (larva). After

140 120 100 80 15 % Poly C 60 VINCRISTINE 40 20 0 CCRF HL-60 K-562 RPMI-8226 SR Cell lines

Comparision for anticancer activity of Polycarponin C with standard Vincristine

Graph. 2: Comparison of synthesized Polycarponin C for anticancer screening with data of vincristine against leukemia human tumor cell lines.

Human Tumor Cell Line	% GI for comp. VIII	Human Tumor Cell Line	% GI for comp. VIII	
Leukemia		Ovarian Cancer		
CCRF-CEM	15.33	IGROV1	13.6	
HL-60(TB)	22.99			
K-562	39.17			
RPMI-8226	15.09			
SR	61.87			
Colon cancer		Renal Cancer		
HCT-116	12.39	UO-31	15.89	
HCT-15	11.79			
HT29	7.01			
KM12	13.9			
SW-620	8.69			
CNS Cancer		Breast Cancer		
SNB-75	18.20	MCF7	31.78	
Melanoma		Mean	90.55	
LOX IMVI	5.59	Delta	61.87	
MALME-3	12.89	ange	98.09	
MDA-MB-435	26.76			

# Table 2: %GI shown by Polycarponin C against different human tumor cell lines

two days, when the shrimp larvae are ready, 4 ml of the artificial seawater was added to each test tube containing different conc. of drug and 10 brine shrimps were introduced into each tube. Thus, there were a total of 30 shrimps per dilution. Then the volume was adjusted with artificial seawater up to 5 ml per test tube. The test tubes were left uncovered under the lamp. The number of surviving shrimps were counted and recorded after 24 hours. The lethality concentration ( $LC_{50}$ ) was assessed at 95% confidence intervals. The percentage mortality

(%M) was also calculated by dividing the number of dead nauplii by the total number, and then multiplied by 100%. This is to ensure that the death (mortality) of the nauplii is attributed to the activity of the compound. The results of activity are shown in Table 1.

# In vitro cytotoxic activity against Human tumor cell lines<sup>12</sup>

The synthesized compoun dwas screened for in vitro anticancer assay by National Cancer



Where: a= DIPC, NMM, CHCl3, RT, 24h, b= TFA, NMM,RT,1h, c= LiOH, THF:H2O(1:1), reflux, 15 mins d= pnp-, CHCl3, RT, 12h, e= NMM, CHCl3, 0°C, 7days Scheme 1: Synthetic route for Polycarponin C

Institute (NCI), Bethesda, USA in a panel of 60 human tumor cell lines. The screening of the compound operated with In Vitro Cell Line Screening Project (IVCLSP), which is dedicated service, providing direct support to the DTP anticancer drug discovery program. The screening was carried out against 60 different human tumor cell lines of the leukemia, Non-smallcell lung, colon, CNS, melanoma, ovarian, renal, Prostrate and breast cancers which was aimed in showing selective growth inhibition or cell killing of particular tumor cell lines by specific compound. The screening begins with the evaluation of selected compounds at asingle dose of 10<sup>-5</sup> M.The output from the single dose screen is reported as a mean graph and is available for analysis by the compare program. The result of anticancer screening is shown in fig. 1 and Table 2.

### **RESULTS AND DISCUSSION**

## Spectral characterization Physical state: Semisolid mass

IR data: intense C=O absorptions at 3300 and 1650 cm<sup>-1</sup>. absorptions at 3427 , 1650 cm<sup>-1</sup>, 1650 cm<sup>-1</sup> are due to amino and amide carbonyl groups respectively FABMS showed M<sup>+</sup> ion peak at m/z 860.

### 13C NMR: showed six amide carbonyl carbon

 $\delta$  171 for C=O of Thr,  $\delta$  62.5 for Cα of Thr,  $\delta$ 66.5 for Cβ of Thr,  $\delta$  22 for Cλ of Thr,  $\delta$  172 for C=O of Phe, $\delta$  52.9 for Cα of Phe,  $\delta$  38 for Cβ of Phe,  $\delta$ 171.5 for C=O of Val,  $\delta$  62 for Cα of Val,  $\delta$  31.5 for Cβ of Val,  $\delta$  19 for Cλ of Val,  $\delta$  170.5 for C=O of Leu,  $\alpha$  50 for Cα of Leu,  $\delta$  41.5 for C $\delta$  of Leu,  $\delta$  25.1 for C $\tilde{\alpha}$  of Leu,  $\delta$  172.5 for C=O of Leu<sub>2</sub>,  $\delta$  51 for C $\beta$  of Leu<sub>2</sub>,  $\delta$  43.5 for C $\beta$  of Leu<sub>2</sub>,  $\delta$  25for Cy of Leu<sub>2</sub>,  $\delta$ 173 for C=O of Pro<sub>1</sub>,  $\delta$  62.2 for C $\alpha$  of Pro<sub>1</sub>,  $\delta$  32.2 for CY of Pro<sub>1</sub>,  $\delta$  22.2 for C $\gamma$  of Pro<sub>1</sub>,  $\delta$  171 for C=O of Pro<sub>2</sub>,  $\delta$  61.2 for C $\hat{\alpha}$  of Pro<sub>2</sub>,  $\delta$  31 for C $\beta$  of Pro<sub>2</sub>,  $\delta$  22

# for C<sub> $\gamma$ </sub> of Pro<sub>2</sub>, ä 170.8 for C=O of Pro<sub>3</sub>, $\delta$ 59.2 for C $\delta$ of Pro<sub>3</sub>, $\delta$ 28.6 for C $\alpha$ of Pro<sub>3</sub>, $\delta$ 25.8 for C $\gamma$ of Pro<sub>3</sub>

#### 1H NMR: showed five amide N-H signals

 $\delta$  7.2 for H<sub>N</sub> of Thr,  $\delta$  4.55 for H<sub>N</sub> of Thr,  $\delta$ 4.65 for Hβ of Thr,  $\delta$  1.36 for Hβ of Thr,  $\delta$  11.04 β for H<sub>N</sub> of Phe,  $\delta$  5.02 for Hα of Phe,  $\delta$  3.01 for Hβ of Phe,  $\delta$  8.7 for H<sub>N</sub> of Val,  $\delta$  4.7 for Hα of Val,  $\delta$ 2.5 for Hβ of Val,  $\delta$  1.5 for Hβ of Val,  $\delta$  8 for H<sub>N</sub> of Leu<sub>1</sub>,  $\delta$  5.36 for Hα of Leu<sub>1</sub>,  $\delta$  1.74 for Hβ of Leu<sub>1</sub>,  $\delta$ 1.90 for Hβ of Leu<sub>1</sub>,  $\delta$  8.5 for H<sub>N</sub> of Leu<sub>2</sub>,  $\delta$  5.05 for Hα of Leu<sub>2</sub>,  $\delta$  1.7 for Hβ of Leu<sub>2</sub>,  $\delta$  1.94 for Hβ of Leu<sub>2</sub>,  $\delta$  4.45 for Hα of Pro<sub>1</sub>,  $\delta$  2.16 for Hβ of Pro<sub>1</sub>,  $\delta$  1.80 r Hβ of Pro<sub>1</sub>,  $\delta$  4.4 for Hα of Pro<sub>2</sub>,  $\delta$  1.90 for Hβ of Pro<sub>2</sub>,  $\delta$  1.65 for Hβ of Pro<sub>2</sub>,  $\delta$  4.39 for Hα of Pro<sub>3</sub>,  $\delta$  1.90 for Hβ of Pro<sub>3</sub>,  $\delta$  2.16 for Hβ of Pro<sub>3</sub>, 4.08 for Hδ of Pro<sub>3</sub>. Elemental analysis: C=63.96, H=8.4, N=13.10, O=14.91

#### CONCLUSION

The compound was synthesized with good yield by using solution phase technique. It showed moderate activity against SR human tumor cell lines of Leukemia when compared with Vincristine as a standard. In consultation with literature survey, synthesis of analogs of this molecule may lead to the development of potent anticancer agents.

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### REFERENCES

4.

- Shinde, N.V.; Himaja, M.; Bhosale, S.K.; Ramana, M.V.; Sakarkar, D.M. Indian J Pharm Sci. 2008, 70(6), 695-852
- Dahiya R.;Gautam H. Bulletin Pharm. Res.2011, 1(1), 1-10
- 3. Chaudhary S.; Kumar H.; Verma H.; Rajpoot

A. Int J. PharmTech Res.2012,4(1), 194-200 Kawagishi H.; Somoto A.;Kuranari J.; Kimura A.; Chiba S. Tetrahedron1993,34(21), 3439-3440

Chaudhary S.;Kumar H.;Verma H.;Rajpoot
 A. Int Journal of PharmTech Res.2012, 4(1),

194-200

- 6. Hernández D. *Eur. J. Org. Chem.***2008**, 3389–3396
- Shinde N.V.;Himaja M.;Bhosale S.K.;Ramana M.V.;Sakarkar D.M. *Asian J Chem.*2010,22(2), 996-1000
- 8. Ding, Z. T.; Zhou, J.; Tan, N.; Cheng, Y.; Deng, S. *ActabotanicaSinica.***2001**, *43(5)*, 541-44
- 9. Bodanszky M. and Bodanszky A., Practice of peptide synthesis;Springer-verlog Publishers. New York, (1984)
- 10. Bussmann R.;Malca G.; Glenn A. J. Ethnopharmacology:**2011**,137, 121-140
- 11. Houghton P.; Fang R.; Techatanawat I. *Science Direct.***2007**, 42, 377–387. www.nci.gov.in

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