Ninhydrin Based Visible Spectrophotometric Determination of Gemigliptin

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ABSTRACT

A simple method is described to determine the amount of gemigliptin in bulk and tablet formulation by visible spectrophotometry. Basis of the proposed method is the reaction of the primary amine present on gemigliptin with ninhydrin in alkaline pH (alkaline borate buffer) medium to produce a purple color (Ruhemann’s purple) which has maximum absorption at 558 nm. The method was validated as per the current ICH guidelines. The obtained regression equation (y = 0.0148x+0.0011) in the range of 5-30 μg mL⁻¹ has a good correlation coefficient (> 0.999). As the method does not require any separation, it is rapid and simple. The recovery levels of the drug were in the range of 99.73 – 99.96. This method is a green method as it no organic solvents were employed.

Keywords: Ruhemann’s purple, Gemigliptin, Ninhydrin, Validation, Determination.

INTRODUCTION

Gemigliptin is a long acting, competitive and potent, selective anti-hyperglycemic agent to treat type 2 diabetes and this drug falls under DPP-4 (dipeptidyl peptidase-4) inhibitor class¹. Levels of incretin hormones level are increased by it which further kindle the secretion of glucose-dependent, depletes the secretion of glucagon, and also helps to decrease postprandial hyperglycemia. It is effective either as in a combination drug with metformin or in monotherapy². Its molecular formula is C₁₈H₁₉F₈N₅O₂ with a molecular weight of 489.36. Its IUPAC name is (3S)-3-amino-4-(5,5-difluoro-2-oxopiperidino)-1-[2,4-di(trifluoromethyl)-5,6,7,8-tetrahydropyrido[3,4-d]pyrimidin-7-yl]butan-1-one (Fig. 1). LG Life Sciences (LGLS) developed it initially. Final product was co-developed with Double-Crane Pharmaceutical Co. and marketed in China. Development and commercialization of it in Turkey were licensed to NOBEL Ilac in November 2010. Zemiglo is the market product of LGLS in Korea. Sanofi (Paris) and Stendhal (Mexico) were permitted by LGLS to market in 104 countries. Other names of it are LC-150444 and Gemiglo®. Its route of administration is oral³.
A thorough literature collection shows that very few methods were proposed to estimate gemigliptin using HPLC-Isocratic and LC/MS-MS. But, no visible spectrophotometric method is available. Chemical structure of gemigliptin shows the presence of amine groups which will be useful for formation of Ruhemann’s purple with ninhydrin. Therefore, the present method describes visible spectrophotometric determination of gemigliptin using ninhydrin.

**MATERIALS AND METHODS**

**Ninhydrin solution**

It was prepared by dissolving 0.2 g of Ninhydrin in 100 mL of buffer (pH 10) solution. On exposure to sunlight, it becomes red. Hence, this solution was refrigerated at 4°C and used for a period of 3 days.

Alkaline borate buffer (pH 10) was prepared as described in Indian Pharmacopoeia.

**Preparation of standard drug solution**

The standard drug of gemigliptin (50 mg) was weighed accurately and transferred to 50 mL volumetric flask. It was dissolved properly and diluted up to the mark with methanol to obtain final concentration of 1000 μg mL⁻¹ (stock solution). This solution was diluted for further suitably.

**RESULTS AND DISCUSSIONS**

Ninhydrin is the chosen reagent in the current study as it reacts with primary aliphatic amines to produce Ruhemann’s purple colour. The absorption spectrum of coloured compound shows λ max as 558 nm (Figure 2).

**Optimization of reactions conditions and stoichiometry**

The optimum conditions for the reaction were found to be 2 mL ninhydrin solution, heating 80±5 °C as heating temperature, buffer solution (pH 10) as diluent. Heating time of the drug and ninhydrin mixture was fixed as 15 min because, maximum absorption was noted for the produced violet colour. Beyond 20 min the colour was unstable and slowly faded out. Different organic solvents (DMF, DMSO, acetone or methanol) were tested to choose the appropriate solvent for diluting the reaction mixture. In addition, water was also tested because the present method does not involve any extraction step before the measurement of absorbance. Moreover, the added advantage is avoiding the usage of toxic organic solvents. Water was found to be the best with a stable and maximum intensity of colour.

In the case of amino acids reaction with ninhydrin, alkaline pH is preferred. Because, the un-protonated amino group behaves as an excellent nucleophile at higher pH, whereas, protonation at lower pH makes it less nucleophilic. However, optimum pH may vary with the nature of amino acid. In the present case, pH 10 is found to be optimum. The noted stoichiometric mole ratio between gemigliptin and ninhydrin is deviating from 1:2 and was towards higher values of ninhydrin for the development of maximum colour intensity. This non-ideal stoichiometry for the Ruhemann’s purple formation in the present study might be due to possible reasons like slower rate of reaction, side reactions, photolytic instability and interferences.
Recommended analytical procedure

Different aliquots of standard gemigliptin solution were transferred into a series of 10 mL boiling test tubes. Ninhydrin (2 mL) was added and the mixture was heated in a water bath at 80 ± 5°C for 15 minutes. The contents were cooled and the volume was made up to the mark with alkaline borate buffer buffer in 10 mL volumetric flasks. The absorbance was measured at 558 nm against a reagent blank.

Chromophore Formation and Chemistry

Ninhydrin is a popular chromogen to determine certain amino acids, amines and thiophenes\(^{13}\). It is a triketohydrindene hydrate and chemically it is 2,2-dihydroxy-1,3-indandione. It is a reagent having carbonyl group and was successfully used for assay of a number of pharmaceutical compounds containing nitrogenous functional groups like amino acids, amines, amides, hydrazines, piperazines and cyanides. Formation of a condensation product having a typical purple colour forms the basis for the spectrophotometric measurement\(^{14}\). It is also known as Ruhemann’s reagent because ninhydrin reacts with primary amino groups to develop a coloured chromophore known as Ruhemann’s purple which is chemically known as diketohydrindylindene-diketohydridamine\(^{15}\).

Faint alkaline condition has to be maintained for reaction of ninhydrin\(^{16}\). Sodium hydroxide or sodium carbonate is generally used for the purpose. Pyridine has the advantage to provide the required alkalinity and maintain reaction conditions\(^{15}\). But a very few researchers reported the reactivity of ninhydrin with primary amine and amides in acidic medium (pH 5.5), in the presence of a catalyst like sodium molybdate to form the same Ruhemann’s purple\(^{14}\). On the other hand, formation of Ruhemann’s blue was reported by Ramadan and Abuinban\(^{17}\) in slightly acidic medium (pH 6.0) in presence of reducing agent- ascorbic acid.

Majority of the publications report the usage of organic solvents (like DMF, DMSO, acetone or methanol) whereas, others used either buffer solution or aqueous solution\(^{14,15}\). As per the new developing trends, new analytical methods need to be green in addition to other best characteristics\(^{16}\). So, one of the experimental objectives of the present study is to avoid the usage of organic solvents. It has a great significance in reduction of analysis cost. Hence, alkaline borate buffer was used as the reaction medium in the present study and no organic solvent was employed.

The actual color of the product varies in light\(^{19}\). \(\lambda_{\text{max}}\) in the present study is 558 nm. Literature survey shows that \(\lambda_{\text{max}}\) depends on the nature of medium. For example, \(\text{I}_{\text{max}}\) was in the range of 595-600 nm when the medium was N,N'-dimethylformamide (DMF)\(^{20-25}\). In buffer solution or aqueous medium, 555 – 571 nm was observed\(^{14,15,26-31}\). This may be due to polar and stearic effects of reactant\(^{32}\). The observed \(\lambda_{\text{max}}\) can be further substantiated from the studies of Mendel Friedman et al.,\(^{33}\) about the effect of solvent nature on absorption maximum of Ruhemann’s purple. According them, \(\lambda_{\text{max}}\) will be around 605 nm in non-aqueous aprotic solvents (like DMSO and DMF) and a hypsochromic shift was observed to 575 nm in non-aqueous protic solvents (formamide) or by the addition of protic solvents to aprotic media due to involvement of hydrogen bonding. Similarly, a shift to 550 nm was observed by the addition of compound capable of forming charge-transfer interactions (pyridine) to non-aqueous aprotic solvents. However, few researchers reported the formation of yellow coloured product having \(\lambda_{\text{max}}\) of 420 nm in saturated NaHCO\(_3\) solution due to reaction between ninhydrin and secondary amine groups present in pharmaceutical drugs like lisinopril\(^{34}\) and terrbutalin sulphate\(^{35}\). Because, primary amine and ammonia produce purple color with ninhydrin and secondary amine produces yellow color complex with Ninhydrin\(^{36}\). In the present case, the formation of Ruhemann’s purple with gemigliptin is shown below (Figure 3).

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![Figure 3. Formation of Ruhemann’s Purple](image-url)
Mechanism involved in the formation of Ruhemann’s purple depends upon the nature of substrate (amino acid, imino acid, primary amine, secondary amine, pyrrole and ammonium salt). In the case of amino acids, ninhydrin undergoes tautomerization to form 1,2,3-indantrione which reacts with amino acid to give Schiff’s base. An aldehyde is yielded by decarboxylation of the formed ketamine along with formation of an intermediate amine which condenses with another ninhydrin molecule to result in the product – Ruhemann’s purple. However, reaction of pharmaceutical drugs with ninhydrin reagent via oxidative deamination of the primary amino group followed by the condensation of the reduced ninhydrin to form the coloured purple reaction product was reported with gabapentin, lisinopril, pregabalin, amlodipine, famotidine. The postulated reaction mechanism involves the essential existence of ninhydrin and as well as its reduced form to form Ruhemann’s purple by amines. Condensation with both forms of ninhydrin proceeds through either simultaneously or sequentially (Figure 4).

Validation of Method

Linearity and range

Linearity was observed in the concentration range of 5 – 30 μg mL$^{-1}$ (Table 1, Fig. 5). The proposed analytical method is linear due to high correlation coefficient ($> 0.999$) for the observed linear regression equation $y = 0.0148x + 0.0011$. Different parameters (optical and regression) are listed out in Table 2.

Table 1: Calibration values of gemigliptin

<table>
<thead>
<tr>
<th>Concentration (μg mL$^{-1}$)</th>
<th>Absorbance*</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>0.0751</td>
</tr>
<tr>
<td>10</td>
<td>0.1494</td>
</tr>
<tr>
<td>15</td>
<td>0.2242</td>
</tr>
<tr>
<td>20</td>
<td>0.2941</td>
</tr>
<tr>
<td>25</td>
<td>0.3651</td>
</tr>
<tr>
<td>30</td>
<td>0.4482</td>
</tr>
</tbody>
</table>

*Average of three determinations

Accuracy

Percent recovery values were determined in order to ascertain the accuracy of the proposed method. Various quantities of gemigliptin (5, 10 and 15 μg mL$^{-1}$) were added to a nominal amount (10 μg mL$^{-1}$) to study recovery levels of 50%, 100% and 150%
respectively. Range of % recovery values is 99.73 – 99.96 as observed from Table 3. Acceptable level of accuracy is demonstrated from low values of SD and %RSD.

Table 2: Optical, regression and validation parameter values

<table>
<thead>
<tr>
<th>S. No.</th>
<th>Parameter</th>
<th>Observation</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Apparent molar absorptivity (L mol⁻¹ cm⁻¹)</td>
<td>2.53×10⁴</td>
</tr>
<tr>
<td>2</td>
<td>Sandell’s sensitivity (μg cm⁻²A⁻¹)</td>
<td>0.0673</td>
</tr>
<tr>
<td>1</td>
<td>Slope</td>
<td>0.0148</td>
</tr>
<tr>
<td>2</td>
<td>Intercept</td>
<td>0.0011</td>
</tr>
<tr>
<td>3</td>
<td>Regression coefficient (r)</td>
<td>0.9995</td>
</tr>
</tbody>
</table>

Table 3: Recovery of gemigliptin

<table>
<thead>
<tr>
<th>Level of recovery(%)</th>
<th>Amount of drug (Practical) recovered (μg mL⁻¹)</th>
<th>Statistical evaluation % Recovered</th>
<th>Recovery = Practical x 100/ Theoretical</th>
</tr>
</thead>
<tbody>
<tr>
<td>50</td>
<td>14.96 Mean</td>
<td>14.98</td>
<td>99.73</td>
</tr>
<tr>
<td></td>
<td>14.98 SD</td>
<td>0.013</td>
<td>99.86</td>
</tr>
<tr>
<td></td>
<td>14.99 %RSD</td>
<td>0.083</td>
<td>99.93</td>
</tr>
<tr>
<td>100</td>
<td>19.96 Mean</td>
<td>19.97</td>
<td>99.80</td>
</tr>
<tr>
<td></td>
<td>19.98 SD</td>
<td>0.008</td>
<td>99.90</td>
</tr>
<tr>
<td></td>
<td>19.97 %RSD</td>
<td>0.040</td>
<td>99.85</td>
</tr>
<tr>
<td>150</td>
<td>24.98 Mean</td>
<td>24.98</td>
<td>99.92</td>
</tr>
<tr>
<td></td>
<td>24.99 SD</td>
<td>0.008</td>
<td>99.96</td>
</tr>
<tr>
<td></td>
<td>24.97 %RSD</td>
<td>0.032</td>
<td>99.88</td>
</tr>
</tbody>
</table>

Table 4: Precision data

<table>
<thead>
<tr>
<th>Concentration*</th>
<th>Intraday %RSD (Mean ± SD)</th>
<th>Inter-day %RSD (Mean ± SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>5.003±0.026 0.520</td>
<td>5.007±0.048 0.959</td>
</tr>
<tr>
<td>15</td>
<td>15.174±0.048 0.316</td>
<td>15.241±0.052 0.341</td>
</tr>
<tr>
<td>30</td>
<td>30.058±0.051 0.170</td>
<td>30.084±0.058 0.193</td>
</tr>
</tbody>
</table>

*Average of six determinations

Detection limits determination

Both the limits for quantification as well as detection were calculated using the values of signal to noise ratioLOQ as well as LOD for gemigliptin determination using the current method was done using values of S (calibration curve slope) and σ (response standard deviation)

LOD = 3.3 × σ/S = 0.15 μg mL⁻¹
LOQ = 10 × σ/S = 0.50 μg mL⁻¹
Pharmaceutical Formulations Analysis

Sonnicated a mixture of methanol and tablet formulation (ZEMIGLO®) powder for ten minutes to extract the API present in it. Then the amount of API was determined by adopting the above method (Table 6). Visible spectrophotometric method is the generally used for routine analysis in industrial QC laboratories, especially in developing countries13-49. Hence, the developed visible spectrophotometric method can be adopted for the determination of gemigliptin in pure and tablet formulations using Ninhydrin as a chromogenic agent.

Table 6: Assay of Pharmaceutical Formulation

<table>
<thead>
<tr>
<th>Formulation</th>
<th>Labeled amount (g)</th>
<th>Amount found* (g)</th>
<th>% Drug Recovered</th>
<th>%RSD</th>
</tr>
</thead>
<tbody>
<tr>
<td>ZEMIGLO®</td>
<td>50</td>
<td>50.0214±0.0028</td>
<td>100.04</td>
<td>0.006</td>
</tr>
</tbody>
</table>

* Average of three determinations

CONCLUSION

Formation of a purple color (Ruhemann’s purple) is due to the reaction between primary amine present on gemigliptin and ninhydrin in alkaline pH medium. As organic solvents are avoided in the present method, it can be considered as a green method. The proposed method was validated as per the existing guidelines of ICH. As alternative to the expensive instrumental methods, this method can be considered for routine analysis of gemigliptin (bulk drug and tablet formulation) in quality control laboratories.

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