INTRODUCTION

Generally, bioethanol converted from edible sources is called first generation bioethanol (FGB). However, the drawback of FGB is that it stems from edible feed stock utilized, which include corn and sugarcane\(^1\). The technology development focuses on the production of ethanol has shifted towards the utilization of residual lignocellulosic materials to lower production costs\(^2\). The enzyme then converting the sugar released into ethanol through the metabolism of \textit{Saccharomyces cerevisiae}\(^3\).

Corn stalk have high content of easily hydrolysable polysaccharides called pentosic. Compositions of the corn stalk are also obtained from the analysis, where it consists of 19.35\% lignin, 40.28\% Cellulose, 35.06\% Pentosans\(^3\). The use of corn stalk as sources for ethanol can contribute in low cost production of ethanol.

SYNTHESIS OF BIO-ETHANOL FROM CORN STALK BY FERMENTATION PROCESS

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ABSTRACT

Ethanol synthesis from corn stalk through fermentation process was studied. The ethanol produced was studied by various enzymatic treatment temperature and different feedstock loading in term of weight. The result shows that the highest concentration of ethanol contained in the sample was 48.90\% at enzymatic temperature of 50\(^\circ\)C. The temperature for optimum enzyme treatment have been identified as 50\(^\circ\)C followed by 30, 40 and 60\(^\circ\)C respectively.

Key words: Ethanol, Bio-ethanol, Enzyme treatment, Enzymatic temperature.
There is certainly industry addressing the cellulosic ethanol sector by making the process cheaper and more competitive with ethanol produced from sugar and starch sources\(^4\).

Objectives for this study are to analyse the composition of ethanol produced from the process using High Performance Liquid Chromatography (HPLC) and to study the production of bioethanol from corn stalks through enzymatic hydrolysis.

**MATERIALS AND METHOD**

**Preparation of Corn Stalks**

In this experiment, the corn stalk were obtained from Pasir Mas, Kelantan. The corn stalk obtained are from matured stalk that have been harvested for the corn. After obtaining the corn stalk, the leaves were removed and then dried at the temperature of 60-70°C until constant weight was achieved. The dried corn stalk then were chopped manually into small pieces of 10-30 mm. Finally the corn stalk were grinded into smaller pieces of 0.2-2 mm ready for the enzyme treatment.

**Enzyme treatment**

The grinded corn stalk were slurried with acetate buffer (0.05M, pH 4.8) at 5% (w/v) substrate loading then were autoclaved at 121°C for 15 minutes\(^5\). After autoclaved, the sample was cooled to 50°C, 2% Tween 20 (v/v) was then loaded into the sample as surfactant. The sample mixture were treated with 0.3% (v/v) Viscozyme® L Cellulolytic Enzyme Mixture V2010 from Sigma Aldrich and was incubated on an orbital shaker 150rpm at different temperatures of 30°, 40°, 50° and 60° C for 48 hours. For each temperature, the sample have 10, 20 and 30 gram of corn stalk loading.

**Microorganisms and batch fermentation**

Commercial baker's yeast, Mauripan were used as the source of *Saccharomyces cerevisiae*. The dry yeast were inoculated into the medium consists of: glucose 50g/L, peptone 5g/L, MgSO\(_4\), 7H\(_2\)O 1g/L, K\(_2\)HPO\(_4\) and 5g/L of Yeast. The medium was autoclaved at 121 p C for 15 minutes. After autoclaved, the yeast was inoculated on the orbital shaker for a period of 18 hours at 30°C, 50rpm.

The media containing yeast then was added at the volume ratio of 1:10 of the fermentation broth aseptically. The fermentation of all samples were conducted at 30.5 p C at 150rpm for a period of 48 hours using an orbital shaker\(^6\).

**RESULTS AND DISCUSSION**

**High Performance Liquid Chromatorgraphy (HPLC) Analysis**

1.5 ml of samples after rotary evaporator were loaded into the HPLC vial as sample preparation. 20µl was injected into the HPLC system to analyse the presence of ethanol in the sample. The HPLC analysis parameter were determined using the following conditions: column, C18 RP Hyper Sil; 20µl of sample was injected into the HPLC system. The mobile phase was 0.1M pH 2.5 Phosphoric Acid and the flow rate was 1.5ml/min and the detection was set at a wavelength of 254nm\(^7\).
for corn stalk loading of 30 gram. Then followed by 20 gram at 9629 and lastly 10 gram peak area of 2437. The peak area produced then were plotted into Figure 2 and the value of R²=0.9942 shows close correlation between the peak area and the corn stalk weight. Ethanol concentration were then calculated by substituting the peak area obtained from the HPLC analysis into y of the equation 1. The ethanol concentration produced then were plotted into linear graph (Figure 3). Ethanol concentration produce shows increasing trend with the increase of corn stalk weight. The highest concentration were 48.90% for corn stalk loading of 30 gram followed by 33.92% at 20 gram and 14.38% for 10 gram of corn stalk loading.

Table 3 shows the mean peak area for enzyme treatment at 30°C with the highest area were 8304.5 for corn stalk weight 30 gram and the lowest was 10 gram corn stalk loading with peak concentration 14.38%.

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Table 1: Absolute ethanol reading on the HPLC

<table>
<thead>
<tr>
<th>Ethanol Concentration (%)</th>
<th>Retention time (min)</th>
<th>Peak Area</th>
<th>Area (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>100</td>
<td>2.856</td>
<td>31,497</td>
<td>89.328</td>
</tr>
<tr>
<td>75</td>
<td>2.886</td>
<td>28,520</td>
<td>91.681</td>
</tr>
<tr>
<td>50</td>
<td>2.940</td>
<td>15,330</td>
<td>3.528</td>
</tr>
<tr>
<td>25</td>
<td>3.145</td>
<td>5229</td>
<td>1.685</td>
</tr>
</tbody>
</table>

Table 2: Mean retention time and peak area for sample done at temperature of 50°C

<table>
<thead>
<tr>
<th>Sample at different weight (g)</th>
<th>Mean retention time (min)</th>
<th>Mean peak area</th>
<th>Mean peak height</th>
</tr>
</thead>
<tbody>
<tr>
<td>10</td>
<td>3.573</td>
<td>2437</td>
<td>420</td>
</tr>
<tr>
<td>20</td>
<td>3.478</td>
<td>9629</td>
<td>1430</td>
</tr>
<tr>
<td>30</td>
<td>3.118</td>
<td>15142.5</td>
<td>1571.5</td>
</tr>
</tbody>
</table>

Table 3: Mean retention time and peak area for sample done at temperature of 30°C

<table>
<thead>
<tr>
<th>Sample at different weight (g)</th>
<th>Mean retention time (min)</th>
<th>Mean peak area</th>
<th>Mean peak height</th>
</tr>
</thead>
<tbody>
<tr>
<td>10</td>
<td>3.568</td>
<td>3340.5</td>
<td>477.5</td>
</tr>
<tr>
<td>20</td>
<td>3.523</td>
<td>5740.5</td>
<td>803.5</td>
</tr>
<tr>
<td>30</td>
<td>3.531</td>
<td>8304.5</td>
<td>1073</td>
</tr>
</tbody>
</table>

Table 4: Mean retention time and peak area for sample done at temperature of 40°C

<table>
<thead>
<tr>
<th>Sample at different weight (g)</th>
<th>Retention time (min)</th>
<th>Peak area</th>
<th>Peak height</th>
</tr>
</thead>
<tbody>
<tr>
<td>10</td>
<td>2.215</td>
<td>5525</td>
<td>1109</td>
</tr>
<tr>
<td>20</td>
<td>2.147</td>
<td>1772</td>
<td>311</td>
</tr>
<tr>
<td>30</td>
<td>N/A</td>
<td>N/A</td>
<td>N/A</td>
</tr>
</tbody>
</table>
Fig. 1: Linear graph plot of ethanol calibration

Fig. 2: Graph plot of the peak area for sample treatment at 50°C

Fig. 3: Graph plot of the ethanol concentration of sample treatment at 50°C
Fig. 4: Graph plot of the peak area for sample treatment at 30°C

Fig. 5: Graph plot of the ethanol concentration of sample treatment at 30°C

Fig. 6: Graph plot of the ethanol concentration of sample treatment at 40°C
area of 3340.5. Figure 4 shows the interaction between the peak area of 10, 20 and 30 gram of corn stalk, the straight line produce value of $R^2=0.9996$ meaning that the all the points correlate between each other. Peak area value were then used to calculate the ethanol concentration of ethanol presence in the sample. Figure 5 shows the straight line produced from the ethanol concentration of sample at different weight. The ethanol increased with the increase of corn stalk weight, the highest ethanol concentration for 30°C enzyme treatment is 30.33% and the lowest is 16.83% ethanol presence on the sample.

Comparing the ethanol concentration between enzyme treatment at 30 and at 50°C, treatment at 50°C produce the highest ethanol concentration than treatment at 30°C, the highest concentration of 48.90% rather than 30.33% shows large difference between these two concentration. For 20 gram of corn stalk 50°C enzyme treatment also produce higher ethanol concentration at 33.92% rather than 23.36% at 30°C.

Treatment of 40°C during enzyme treatment produce a deviation in the ethanol concentration. Table 4 shows the peak area reading of the sample enzyme treatment at temperature 40°C, Figure 6 shows the linear graph of the ethanol concentration produced, the concentration produce a negative gradient declination from 22.77% of 10 gram of corn stalk to 12.57% (20gram) and no ethanol produced for 30 gram, this due to the oxidization of ethanol to acetic acid result from contamination from microorganisms probably Acetobacter sp. Have been seeing in converting these ethanol into acetic acid in most biological process.

Finally, enzyme treatment at 60°C did not yield any ethanol for any of the corn stalk loading. This is due to the fact that the temperature exceed the optimum temperature of the enzyme used to convert the lignocellulose materials. Thus did not show any peak during the HPLC analysis due to the fermentation process unable to convert any fermentable sugar on the samples.

**CONCLUSION**

From the results produced, it can be concluded that the production of ethanol from corn stalk through enzymatic hydrolysis have been successful. However, the quantity of the production is still small with only the highest concentration is 48.90%. Temperature of the enzyme hydrolysis have been determined to effect on the production of ethanol with 50°C have been the optimum temperature in producing higher ethanol yield for corn stalk. Further research need to be done to determine the optimum condition in producing ethanol and thus producing higher concentration of ethanol for corn stalk as feedstock materials.

**ACKNOWLEDGMENTS**

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**REFERENCES**